

CENTRAL VALLEY REGIONAL WATER QUALITY CONTROL BOARD  
CENTRAL VALLEY REGION

MONITORING AND REPORTING PROGRAM  
ORDER NO. R5-2009-0809  
FOR  
CALIFORNIA RICE COMMISSION

UNDER RESOLUTION NO. R5-2006-0053  
AS AMENDED BY R5-2006-0077  
CONDITIONAL WAIVER OF  
WASTE DISCHARGE REQUIREMENTS  
FOR  
DISCHARGES FROM IRRIGATED LANDS

This Monitoring and Reporting Program (MRP) Order is issued pursuant to the California Water Code (Water Code) sections 13267 and 13269 which authorize the California Regional Water Quality Control Board (hereafter Central Valley Water Board) to require preparation and submittal of technical and monitoring reports. Water Code section 13269 requires a waiver of waste discharge requirements to include as a condition the performance of monitoring and the public availability of monitoring results.

The Executive Officer is issuing this MRP Order to establish specific monitoring and reporting requirements for the California Rice Commission (CRC). The CRC is enrolled under *Amended Coalition Group Conditional Waiver of Waste Discharge Requirements for Discharges from Irrigated Lands, Order No. R5-2006-0053 (Waiver)*. The CRC represents individual rice growers in the Sacramento Valley and covers approximately 500,000 acres.

This MRP Order meets or exceeds the minimum requirements of the Waiver. The MRP Order is additionally consistent with Monitoring and Reporting Program Order No. R5-2008-0005 for Coalition Groups Under Amended Order No. R5-2006-0053 (Coalition Group MRP Order) and meets or exceeds the monitoring and reporting requirements under the Coalition Group MRP Order. This Order rescinds MRP Order No. R5-2007-0835 and shall remain in effect until **31 December 2009**, or when replaced by a revised MRP Order approved by the Executive Officer. The Waiver and other evidence supporting issuing this MRP Order can be found in the Central Valley Water Board's public files. The Information Sheet for the CRC MRP Order (Attachment A), which identifies the regulatory background, program objectives, and development of minimum requirements, is incorporated as part of this Order.

Pursuant to Water Code section 13269(a)(2), monitoring requirements must be designed to support the development and implementation of the waiver program, including, but not limited to, verifying the adequacy and effectiveness of the Waiver's conditions. The reports required by this MRP Order are needed to evaluate impacts of

discharges of waste from irrigated agricultural operations to waters of the state, to determine compliance with the Waiver, and to support the development and implementation of the Waiver as it applies to the CRC Coalition and its members. As provided in the Waiver, this MRP Order is issued to the CRC Coalition, because the CRC Coalition represents irrigated agricultural facilities that discharge waste to waters of the State.

The submittal of an acceptable Quality Assurance Program Plan (QAPP) that meets the requirements of this MRP Order is a condition of the Waiver. The QAPP must be submitted to the Central Valley Water Board within two months of the adoption of this MRP Order, or as directed by the Executive Officer.

The CRC has been monitoring water quality and reporting monitoring results to the Central Valley Water Board's Irrigated Lands Regulatory Program since September 2004. The results indicate that water quality is impacted by unknown source(s) for aquatic algae toxicity and the Executive Officer has requested the preparation of a Management Plan to address the impact. The Management Plan sites are identified as Special Project monitoring sites.

The timing of the submittal process is further clarified as follows:

ACTION	ACTION DEADLINE
Submittal of CRC QAPP	Two months from the adoption of MRP Order
Revised CRC Management Plan for algae toxicity	Two months from the adoption of MRP Order
Submittal of monitoring results from UCD grant*	31 March 2009

\* All available UC Davis grant monitoring results, including TOC, DOC, nitrate, phosphate. The UCD grant is described in section IV.D.

### **PART I. MRP ORDER OBJECTIVES**

The Water Code mandates that monitoring requirements for a Waiver be designed to verify the adequacy and effectiveness of the Waiver's conditions. One of the conditions of the Waiver is that discharges of waste from irrigated lands to surface waters of the State shall not cause or contribute to an exceedance of an applicable water quality standard. Water quality standards are defined for the Irrigated Lands Regulatory Program (ILRP) in the Waiver for the Coalition Group and in Attachment B (Applicable Definitions and Acronyms) of this Order.

The objectives for a MRP Plan are identified in the attached Attachment A -- Information Sheet, which is part of this MRP Order. Implementation of this Order must provide information to determine whether discharges are in compliance with the conditions of the Waiver, including compliance with applicable water quality standards.

## PART II. COMPONENTS

### A. MONITORING SITE INFORMATION

The monitoring strategy for this MRP Order includes the different types of monitoring needed to meet MRP Order objectives. These include Assessment Monitoring for the condition of the water body and Core Monitoring for trends. The Core Monitoring sites identified in this MRP Order are sites previously monitored by the CRC for the last four years. Assessment Monitoring sites are new sites that will provide data on water bodies representing a range of hydrologic conditions, provide data to develop correlations between Assessment sites and Core sites, provide upstream data on new generation pesticides, and monitor water quality from drainages with a high percentage of land farmed in rice. The monitoring sites are sufficiently representative to generally characterize water quality for surface waters of the State that may be affected by rice discharges. The sites are identified in Table 1 below.

<b>Table 1. Monitoring Sites</b>		
<b><u>Site Code</u></b>	<b><u>Site Name*</u></b>	<b><u>Type of Monitoring</u></b>
CBD5	Colusa Basin Drain #5	Core & Special Monitoring
BS1	Butte Slough at Lower Pass Rd	Core & Special Monitoring
CBD1	Colusa Basin Drain above Knights Landing	Core & Special Monitoring
SSB	Sacramento Slough Bridge near Karnak	Core & Special Monitoring
F	Lurline Creek; upstream site of CBD5	Assessment
G	Cherokee Canal, upstream site for BS1**	Assessment
H	Obanion Outfall at DWR PP on Obanion Rd	Assessment

\* Latitude and Longitude provided in Order Attachment D

\*\* If there is no flow at the specified site, a site on Butte Slough will be sampled.

The four core sites have Irrigated Lands Regulatory Program monitoring data for four years and will be used to track trends. The three assessment sites are newly established monitoring sites and will be used to substantiate the representative

reliability of the core sites. Any constituent in the assessment monitoring parameters not previously analyzed at the core sites will be analyzed for at least one year.

### **B. QUALITY ASSURANCE PROJECT PLAN (QAPP)**

The CRC must develop a QAPP to include watershed and site-specific information, project organization and responsibilities, and the quality assurance components of Attachment C of this MRP Order. Attachment C presents the requirements and the guidelines for development of the CRC QAPP, including the laboratory and field requirements to be used for data evaluation. The Central Valley Water Board may conduct an audit of the CRC's contracted laboratories at any time in order to evaluate compliance with the QAPP. Quality control requirements are applicable to all the constituents listed in the Attachment C, as described in the appropriate method.

### **C. MANAGEMENT PLAN FOR ALGAE TOXICITY**

The Management Plan for Algae Toxicity (Management Plan) is required as Special Project Monitoring, described in Attachment A. The Management Plan was conditionally approved on 9 June 2008 and must be updated with the findings from recent monitoring.

## **PART III. MONITORING STRATEGY AND SCHEDULE**

### **A. MONITORING SCHEDULE**

Monitoring periods for CRC sampling are based on the timing and frequency of discharge from rice fields that may contain constituents that affect water quality. For the purposes of this MRP Order, the irrigation monitoring season is defined as April through August with some variance due to the effects of weather on rice activities. Peak pesticide use on rice occurs during the months of April through June. The greatest risk to water quality and the likelihood of rice pesticide detections also occurs during the peak application period.

Prior to harvest, rice fields are drained en masse. The rice drainage season is typically mid-August through September. For decomposition of rice straw and to provide waterfowl habitat, rice fields are generally flooded. The fields are drained from mid-February to March and prepared for the next season. No applications of fertilizers or pesticides occur on rice fields during the winter until the fields are drained.

Monitoring shall be conducted as shown in Table 2. Factors, such as weather conditions, may affect planting and pesticide application. Timing of monitoring will take into account these factors and may vary from year to year. Due to the unique schedule for rice activities, reporting of monitoring results will occur annually.

<b>Table 2. Summary of Assessment and Core Monitoring Periods</b>				
<b>Rice Farming Calendar</b>		<b>Month</b>	<b>Assessment sites</b>	<b>Core sites</b>
Winter drainage		mid-February thru March	No monitoring	No monitoring
Irrigation season	Peak Pesticide Use Season	April thru June	4 times during irrigation season	4 times during irrigation season
		July thru mid-August		
Fall drainage		mid-August thru September	2 times	2 times
		Late September	Sediment sampling 1 time	
Winter flood		October thru November	No monitoring	No monitoring

**B. MONITORING PARAMETERS**

Water quality and flow monitoring shall be used to assess the wastes in discharges from rice fields to surface waters and to evaluate the effectiveness of management practice implementation efforts. Water quality is evaluated by both field-measured parameters and laboratory analytical data. Field measured parameters shall include flow, pH, electrical conductivity, temperature, and dissolved oxygen. Laboratory analytical data must include, the list of constituents, parameters, and tests in Table 3 of this MRP Order. This table identifies the constituents to be analyzed for Assessment and Core monitoring. Site conditions shall be documented by taking digital photos at every monitoring site during each monitoring event.

Acceptable methods for laboratory field procedures as well as quantitation limits are described in Attachment C. Quality control requirements are applicable to all the constituents in Attachment C, as listed in the appropriate method. Analytical procedures that are not conducted under methods listed in the Attachment C, or for analytes that are not listed in the methods in Attachment C, must follow EPA protocols for performance-based method validation.<sup>1</sup>

<sup>1</sup> USEPA, Protocol for EPA Approval of Alternate Test Procedures for Organic and Inorganic Analytes in Wastewater and Drinking Water, EPA 831-B98-002, March 1999.

<b>Table 3. CRC Monitoring Parameters</b>	
<b>Constituent</b>	<b>Type of Monitoring</b>
<b>General physical parameters</b> Flow <sup>1</sup> pH Electrical conductivity Dissolved oxygen Temperature Hardness Turbidity Total dissolved solids	Assessment and Core
<b>Nutrient Analysis<sup>2</sup></b> Total Kjeldahl nitrogen Nitrate + nitrite, as N Total ammonia Unionized ammonia (calculated) Total phosphorous as P Soluble orthophosphate Total organic carbon (TOC)	To be determined for 2010 – based on UC grant findings
<b>Water column toxicity</b> Selenastrum capricornutum Ceriodaphnia dubia Pimephales promelas	Assessment
<b>Photo monitoring (digital)<sup>3</sup></b>	To be taken initially, and as needed to document site changes that could affect monitoring results
<b>Metals</b> <ul style="list-style-type: none"> <li>• Copper</li> </ul>	Assessment and Core
<b>Pesticides</b> <ul style="list-style-type: none"> <li>• Carfentrazone ethyl</li> <li>• Clomazone</li> <li>• Glyphosate</li> <li>• Pendimethalin</li> <li>• Penoxsulam</li> </ul>	Assessment and Core
<ul style="list-style-type: none"> <li>• Propanil</li> </ul>	To be determined – based on evaluation of registrant information submitted 30 December 2008.

Table 3. CRC Monitoring Parameters	
Constituent	Type of Monitoring
<b>Sediment toxicity</b> Hyallolella azteca	Assessment
<b>Sediment Pesticides</b> Lambda Cyhalothrin S-Cypermethrin	As needed, based on criteria described in Attachment C
<b>Sediment TOC</b>	Assessment

### C. TOXICITY PROCEDURES – TOXICITY IDENTIFICATION EVALUATION (TIE) AND DILUTION SERIES

Discharge to receiving waters and sediment must be evaluated using aquatic toxicity testing. The purpose of the toxicity testing is to: 1) evaluate compliance with the narrative toxicity water quality objective; 2) identify the causes of toxicity when and where it is observed (e.g., metals, pesticides, ammonia, etc.); 3) evaluate any additive toxicity or synergistic effects due to the presence of multiple constituents; and 4) determine the sources of the toxicants identified.

1. WATER COLUMN TOXICITY. Water column toxicity analyses shall be conducted on 100% (undiluted) sample for the initial screening, and sufficient sample volume shall be collected in order to allow the laboratory to conduct a Toxicity Identification Evaluation (TIE) on the same sample, should toxicity be detected, in order to identify the cause of the toxicity. The TIE shall take place immediately if a 50% or greater difference in test organism mortality, as compared to the laboratory control, is detected at any time in an ambient sample during an acceptable *Ceriodaphnia dubia* or *Pimephales promelas* test. A TIE shall also be initiated immediately if a 50% or greater reduction in test organism growth is detected between an ambient sample and the laboratory control at the end of an acceptable *Selenastrum capricornutum* test. The frequency of conducting TIEs for *Selenastrum capricornutum* tests may be modified by an approved management plan. At a minimum, Phase I TIE<sup>2</sup> manipulations shall be conducted to determine the general class (e.g., metals, non-polar organics, and polar organics) of the chemical causing toxicity. Phase II<sup>3</sup> TIEs may also be utilized to confirm and identify specific toxic agents. The TIE report to the Regional Central Valley Water Board must include a detailed description of the specific TIE manipulations that were utilized (Section B.5.5 of Attachment C).

<sup>2</sup> USEPA. 1998. Methods for Aquatic Toxicity Identification Evaluations. Phase I Toxicity Characterization Procedures. Office of Research and Development, Duluth, MN. EPA-600-3-88-034.

<sup>3</sup> USEPA. 1998. Methods for Aquatic Toxicity Identification Evaluations. Phase II Toxicity Identification Procedures. Office of Research and Development, Duluth, MN. EPA-600-3-88-035.

If at any point during the initial toxicity screening the mortality reaches 100%, a multiple dilution test shall be initiated in addition to the TIE. The dilution series must be initiated within 24 hours of the sample reaching 100% mortality, and must include a minimum of five (5) sample dilutions in order to quantify the magnitude of the toxic response.

When a 'statistically significant' reduction is observed for a sample at the end of an acceptable test (i.e. meets the EPA test acceptability criteria), but the magnitude of the reduction between the sample and the control is <20%, follow up field sampling will not be required. Samples that are 'statistically significant' at the end of an acceptable test and that exhibit a  $\geq 20\%$  reduction in organism response compared to the control will require follow-up field sampling. In the absence of changes specified in approved Management Plan(s), the resampling trigger for *Selenastrum capricornutum* shall be  $\geq 20\%$  reduction in organism response compared to the control.

Samples that exhibit a statistically significant reduction in organism response when compared to the laboratory control must still be reported to the Central Valley Water Board as an exceedance of the narrative water quality objective for toxicity.

2. SEDIMENT TOXICITY. Sampling and analysis for sediment toxicity shall be carried out at each location established by this MRP Order. If appropriate sediment (i.e., silt, clay) is present at the site. If appropriate sediment is not present at the designated water quality-monitoring site, an alternative site with appropriate sediment shall be designated for all sediment collection and toxicity testing events. Sediment samples shall be collected and analyzed for toxicity once per year in late September at assessment monitoring sites. The Executive Officer may request different sample collection timing and frequency under a Management Plan. If the CRC wishes to deviate from the written timing and frequency requirements, it must provide written, scientifically defensible justification for the change. This justification must address the intent of the MRP Plan requirement, be scientifically based, and be approved by the Executive Officer.

Sediment samples that show "statistically significant" toxicity to *Hyallela azteca* at the end of an acceptable test and that exhibit a  $\geq 20\%$  reduction in organism survival compared to the control will require pesticide analysis of the same sample in an effort to determine the possible cause of toxicity. When sediment samples are collected for toxicity analysis, additional sample volume sufficient for the recommended chemical and physical analyses must be collected. This additional sample volume must be held in frozen storage until the results of the toxicity analysis are available. If the sample is not toxic to the test species, the additional sample volume can be discarded.

All sediment samples for Assessment Monitoring must be analyzed for total organic carbon (TOC). Analysis for TOC is necessary to evaluate the expected magnitude of

toxicity to the test species. If the toxicity criterion described above is exceeded, then the additional sample volume must also be analyzed for lambda-cyhalothrin and cypermethrin, the only two pyrethroids used in rice operations. Analysis at practical reporting limits of 1 ng/g on a dry weight basis for each pesticide is required to allow comparison to established lethal concentrations of these chemicals to the test species. This follow-up analysis must begin within five business days of when the toxicity criterion described above is exceeded.

## **PART IV. REPORTING REQUIREMENTS**

### **A. ANNUAL MONITORING REPORTS**

The annual monitoring report (AMR) shall be submitted by **31 December**, covering monitoring for the 2009 calendar year. The monitoring report shall include the following components:

1. Signed Transmittal Letter;
2. Title page;
3. Table of contents;
4. Executive Summary;
5. Description of the CRC Coalition Group geographical area;
6. Monitoring objectives and design;
7. Sampling site descriptions and rainfall records for the time period covered under the Annual Monitoring Report (AMR);
8. Location map(s) of sampling sites, crops and land uses;
9. Tabulated results of all analyses arranged in tabular form so that the required information is readily discernible (example table is included in (MRP Order Attachment C). Tabulated results shall include all available monitoring data that has been generated from the UC grant, including TOC, DOC and nutrients;
10. Discussion of data to clearly illustrate compliance with the Coalition Group Conditional Waiver, water quality standards, and trigger limits;
11. Electronic data submitted in a SWAMP comparable format;
12. Sampling and analytical methods used;
13. Copy of chain-of-custody forms;
14. Field data sheets, signed laboratory reports, laboratory raw data (as identified in Attachment C);
15. Associated laboratory and field quality control samples results;
16. Summary of Quality Assurance Evaluation results (as identified in Attachment C for Precision, Accuracy and Completeness) ;
17. Specify the method used to obtain flow at each monitoring site during each monitoring event;
18. Electronic or hard copies of photos obtained from all monitoring sites, clearly labeled with site ID and date.

19. Summary of Exceedance Reports submitted during the reporting period and related pesticide use information;
20. Actions taken to address water quality exceedances that have occurred, including but not limited to, revised or additional management practices implemented;
21. Status update on preparation and implementation of all Management Plans and other special projects; and
22. Conclusions and recommendations.

Additional requirements and clarifications necessary for the above annual report components are described below:

**Annual Report Component No. 1—Signed Transmittal Letter**

A transmittal letter shall accompany each report. The transmittal letter shall be signed and contain a penalty of perjury statement by the CRC's authorized agent. This statement shall state:

*"I certify under penalty of law that this document and all attachments were prepared under my direction or supervision in accordance with a system designed to assure that qualified personnel properly gather and evaluate the information submitted. Based on my inquiry of the person or persons who manage the system, or those persons directly responsible for gathering the information, the information submitted is, to the best of my knowledge and belief, true, accurate, and complete. I am aware that there are significant penalties for knowingly submitting false information, including the possibility of fine and imprisonment for violations."*

**Annual Report Component No. 8—Location Maps**

Location map(s) showing the sampling sites, crops, and land uses within the CRC's geographic area must be updated once per year and included in each annual report. An accompanying list or table of monitoring site information must include the site name and identification number, ILRP station code number, and Global Positioning System (GPS) coordinates. The map(s) must contain a level of detail that ensures they are informative and useful. GPS coordinates must be provided as latitude and longitude in the decimal degree coordinate system (at a minimum of five decimal places). The datum must be either WGS 1984 or NAD83, and clearly identified on the map. The source and date of all data layers must be identified on the map(s).

Additionally, the following are recommended as appropriate elements for acceptable map(s):

1. Topographic or shaded relief base map.

2. An appropriate scale for the area to be covered. Examples of some commonly used scales are 1:24,000 (1 inch equals 2,000 feet) and 1:63,360 (1 inch equals 5,280 feet).
3. Rural roads, highways, interstates, and railways, as well as city and town names and principal roadways, shown and clearly labeled down to the limits of the map scale.
4. All natural and constructed waterways (including lakes, rivers and irrigation canals) shown and clearly labeled to the limits of the map scale. Flow direction should be indicated.
5. Special features (e.g., weirs, turnouts, operational spill locations, gauging stations, reservoirs, and ponding basins) clearly marked and identified by name.
6. An electronic copy of all data layers created by the CRC (e.g., rice crop boundaries and monitoring sites) in a GIS usable format (e.g., shapefile or geodatabase).

**Annual Report Component No. 9 – Tabulated results**

Data shall be reported in tabular form so that the required information is readily discernible. The data shall be summarized in such a manner to clearly illustrate compliance with the Coalition Group Conditional Waiver.

**Annual Report Component No. 10—Data Discussion to Illustrate Compliance**

The annual report shall include a discussion of the year's data to illustrate compliance with the Coalition Group Conditional Waiver. If a required component was not met, an explanation for the missing data must be included. Results must also be compared to water quality standards and trigger limits.

**Annual Report Component No.11—Electronic Data Submittal**

Electronic submittal of the field and laboratory data in a SWAMP comparable format must be included with the AMR. Exceptions to the due date for submittal of electronic data may be granted by the Executive Officer if sufficient rationale exists.

Electronic data packages are to be submitted to the Central Valley Water Board in accordance with one of the two options, and the method that the Coalition Group elects to utilize must be identified in their MRP Plan. These two options are described below:

**OPTION A. ELECTRONIC SUBMITTAL DATA PACKAGE IN A SPREADSHEET FORMAT**

Under this option all laboratory data must be entered and submitted within the ILRP SWAMP comparable data spreadsheets (EXCEL, or similar spreadsheet) provided by the Central Valley Water Board staff. Under this option, field data will not be required to be submitted electronically. However, in exchange, the CRC will be required to use and complete ILRP SWAMP comparable field sheets (paper copy)

as well as the required spreadsheets for submittal of laboratory data.

The completed required SWAMP comparable field sheets must be included within the AMR if the CRC elects to utilize Option A.

#### OPTION B. ELECTRONIC SUBMITTAL DATA PACKAGE IN A SWAMP DATABASE FORMAT

Under this option all field and laboratory data must be uploaded into a SWAMP comparable database. The CRC will manage this database and all data entry or upload. The CRC will need to work closely with the Central Valley Water Board staff and the SWAMP program to ensure that the database architecture is kept up-to-date and comparable.

Data submitted must be SWAMP comparable in a content and format that is consistent with the requirements of the ILRP. Data that is considered SWAMP comparable must meet the following conditions:

1. Electronic data must be formatted and follow the specifications in the most current *Required Data Submission Format* posted on the ILRP website. This document will be updated on a regular basis to ensure comparability with the SWAMP Program.
2. In addition to the field sample results for laboratory analyses, the content of the submittals must include field and laboratory quality control results as prescribed within the Attachment C, including but not limited to spike analyses, blanks, surrogates and certified reference materials, if applicable.
3. For toxicity analyses, the content of electronic data submittals must include the following:
  - Individual sample results
  - Negative control summary results
  - Replicate results
4. For toxicity analyses, the minimum water quality measurements performed on the test water shall include: electrical conductivity, pH, Ammonia, Temperature, and Dissolved Oxygen. The timing and frequency of these measurements will be determined by the method. If daily measurements are taken then the minimum and maximum measurements of the range must be reported
5. Data that does not meet the project quality assurance acceptance guidelines must be flagged accordingly and must include brief notes detailing the problem within the provided comments field.

Prior to submittal, the data shall be reviewed by the CRC and determined to the best of their knowledge to be free of errors and in conformance with the project quality assurance acceptance guidelines outlined in the CRC QAPP. The procedures for data entry and data review must follow those outlined in the QAPP.

**Annual Report Components No.13, No.14, and No.15—Copies of Laboratory Reports, Chain-of-Custody Forms and Raw Data.**

Copies of all laboratory analytical reports must be included in the monitoring reports as attachments or provided electronically on a CD. For toxicity reports, all laboratory raw data must be included in the analytical report (including data for failed tests), including copies of all original bench sheets showing the results of individual replicates, such that all calculations and statistics can be reconstructed. For chemistry data, analytical reports must include, at a minimum, the following: a lab narrative describing QC failures, analytical problems and anomalous occurrences; chain of custody (COCs) and sample receipt documentation; all sample results for contract and subcontract laboratories with units, RLs and MDLs; sample preparation, extraction and analysis dates; and results for all QC samples including all field and laboratory blanks, lab control spikes, matrix spikes, field and laboratory duplicates, and surrogate recoveries. Lab raw data such as chromatograms, spectra, summaries of initial and continuing calibrations, sample injection or sequence logs, prep sheets, etc., are not required for submittal, but must be retained for a minimum of five years and be provided to the Central Valley Water Board upon request. All original raw data must be maintained and available for a minimum of five years.

**Annual Report Component No.14—Field Data Sheets**

Copies of all field documentation must be included in the monitoring reports as attachments or provided electronically on a CD. An example of an acceptable field data sheet is provided in Appendix C of MRP Order Attachment C. The monitoring reports need to provide information on field conditions at sampling times including a description of the weather, rainfall, temperature, stream flow, color of the water, odor, and other relevant information that can help in data interpretation.

**Annual Report Component No. 16—Quality Assurance Evaluation (Precision, Accuracy and Completeness)**

A summary of precision and accuracy results (both laboratory and field) is required in the annual monitoring report. The data quality indicators required for the ILRP are identified in MRP Order Attachment C; acceptance criteria for all measurements of precision and accuracy must be identified. The CRC must review all QA/QC results to verify that protocols were followed and identify any results that did not meet acceptance criteria. A summary table or narrative description of all QA/QC results that did not meet objectives must be included in the annual report. Additionally, the report must include a

discussion of how the failed QA/QC results affect the validity of the reported data. The corrective actions to be implemented are described in MRP Order Attachment C.

In addition to precision and accuracy, the CRC must also calculate and report Completeness. Completeness includes the percentage of all quality control results that met acceptance criteria, as well as a determination of project completeness. For further explanation of this requirement, refer to MRP Order Attachment C. Completeness is also defined in MRP Order Attachment B (Applicable Definitions and Acronyms).

The CRC may ask the laboratory to provide assistance with evaluation of their QA/QC data, provided that the Coalition Group prepares the summary table or narrative description of the results for the annual monitoring report.

### **Annual Report Component No. 19—Summary of Exceedance Reports**

A summary of the Exceedance Reports submitted during the monitoring period is required in the AMR. In the event of exceedances for pesticides or toxicity, pesticide use data must be included in the annual monitoring report. Pesticide use information will be acquired from the agricultural commissioner. This requirement is described further in the following section on Exceedance Reports.

### **B. EXCEEDANCE REPORTS**

Exceedances for all parameters shall be reported in the Annual Monitoring Report and within the frequency developed in specific Management Plans. The CRC shall provide exceedance reports if monitoring results show exceedances of water quality standards or trigger limits. When a water quality standard is exceeded at a monitoring location(s), the CRC shall submit an Exceedance Report to the Central Valley Water Board. The estimated flow at the monitoring location and photographs of the site must be included.

The CRC shall evaluate all monitoring data and make a determination of an exceedance no later than five (5) business days after receiving the laboratory analytical report. The Exceedance Report shall be sent by email or fax (916-464-4780) within the next business day, describing the exceedance, the follow-up monitoring, and analysis or other actions the CRC may take to address the exceedance.

When any pesticide or toxicity exceedance is identified, follow-up actions must include an investigation of pesticide use within the watershed area that is physically associated with the exceedance location. This includes all pesticides applied within the area that drains to the monitoring site during the four weeks prior to the exceedance date. The pesticide use information may be acquired from the agricultural commissioner, or from information received from agriculture practitioners within the same drainage area. Results of the pesticide use investigation must be summarized and discussed in the annual monitoring report. The development of an approved Management Plan may

supercede this requirement.

### **C. MANAGEMENT PLANS**

The Executive Officer has requested a Management Plan to address the impact of aquatic algae toxicity and this requirement is incorporated into this Order. Additional Management Plans may be required under this Order if more than one exceedance of the same parameter at the same location occurs within a three-year period. A schedule for Management Plan development and implementation shall be provided to the Central Valley Water Board staff within 10 business days following the occurrence triggering the requirement of the Management Plan. A logical approach to prioritization of Management Plan activities can be addressed in the schedule. The Central Valley Water Board staff will then review the schedule for acceptability and either approve the proposed schedule or require that a different schedule be followed. The CRC may also elect to develop a multi-Coalition Group monitoring effort for a waste constituent that is common to all of the parties involved.

Management Plans must begin with identification of the general type of land-use that is the probable source of the pollutant, such as agriculture, urban, forestry or other. If agriculture can be a source -- in whole or in part -- then further development of the Management Plan as described below is required. If the general land-use source is unknown but could be the result of irrigated agriculture activities, then the Management Plan must develop a study design to eliminate or confirm irrigated agriculture as a source. If a contaminant that is being addressed by the Management Plan can be reasonably assumed through source identification to be caused in whole or in part by irrigated agriculture land use, then additional Management Plan components must include the following:

1. Identification of irrigated agriculture source -- general practice or specific location -- that may be the cause of the water quality problem, or a study design to determine the source.
2. Identification of management practices to be implemented to address the exceedances.
3. Management practice implementation schedule. Implementation may occur through another Water Board regulatory program designed to address the specific exceedances.
4. Management practice performance goals with a schedule.
5. Waste-specific monitoring schedule.
6. A process and schedule for evaluating management practice effectiveness.
7. An identified routine schedule of reporting to the Central Valley Water Board.

Multiple exceedances of different types of contaminants at multiple locations may be prioritized. The prioritization may include considerations such as extent, magnitude and

duration, or be based on a design that assumes that resolution of one type of contaminant (such as sedimentation) may help resolve other types of measured exceedances (such as pesticides, toxicity, DO and pH). The assumptions and prioritizations shall be developed in coordination with the Central Valley Water Board staff, and be included as part of the Management Plan to be approved by the Executive Officer.

Management Plan Reporting must be at least as frequent as that required for the Annual Report monitoring data submittals, and shall provide frequent and sufficient information regarding achievement of the performance goals, and stages when evaluations will occur to determine the effectiveness of the management practice implementation, and if the Management Plan strategies need to be revised.

The CRC shall take affirmative steps to identify appropriate management practices. Such steps may involve conducting management practices workshops and/or developing a management practices worksheet questionnaire to determine the management practices being used in the identified areas. The CRC may conduct such outreach efforts or develop the workshops and worksheets with the assistance of the County Agricultural Commissioners, U.C. Cooperative Extension, Natural Resources Conservation Service, Resource Conservation District, or other appropriate groups or agencies.

At the request of the CRC or upon recommendation by Central Valley Water Board staff, the Executive Officer may provide authorization to exempt the development of a Management Plan if the Executive Officer determines that the exceedance is not likely to be remedied or addressed by a Management Plan.

The Executive Officer may also require the CRC and/or its member Dischargers to develop a Management Plan or to take additional actions if monitoring data or other information indicates that water quality may be jeopardized. The Executive Officer may also increase the monitoring requirements where monitoring results, pesticide use patterns, or other indicators suggest that the increase is warranted.

**D. EDGE OF FIELD MONITORING – UNIVERSITY OF CALIFORNIA, DAVIS (UCD)  
GRANT: DUE 31 MARCH 2009**

UCD was awarded funding by the State Water Resources Control Board on 17 June 2004 (Resolution 2004-0035) to study total organic carbon and dissolved organic carbon (TOC/DOC), total dissolved solids and electrical conductivity (TDS/EC), and turbidity. The grant contract, titled *The Regents of the University of California, University of California Davis - State Water Resources Control Board Grant Agreement No. 04-183-555-0*, specifies reporting requirements.

MONITORING AND REPORTING PROGRAM  
ORDER NO. R5-2009-0809  
FOR CALIFORNIA RICE COMMISSION  
UNDER AMENDED ORDER NO. R5-2006-0053  
COALITION GROUP CONDITIONAL WAIVER OF  
WASTE DISCHARGE REQUIREMENTS  
FOR DISCHARGES FROM IRRIGATED LANDS

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The grant study includes four components designed to assess specific water quality parameters discharged from rice fields cultivated under different conditions. The four components include: (1) monitoring of evaluation of TOC/DOC, TDS/EC, and turbidity outflows from rice fields cultivated under differing straw decomposition and winter flood practices, (2) monitoring of TOC/DOC, TDS/EC, and turbidity in rice field “peripheral drains”, (3) monitoring and assessment to assess the impact of alternative seeding methods on pest management and pesticide outflows from rice fields, and (4) monitoring and assessment to assess the impact of alternative seeding methods on nitrogen and phosphorus outflows from rice fields. Water quality monitoring commenced in April 2006.

The CRC AMR for CY2008 shall include the results of grant monitoring as part of the tabulated monitoring results. The CRC AMR for CY2009 shall include monitoring results, as well as a detailed discussion of the UCD data and any development of management practices that may be implemented or recommended by the CRC for rice fields.

The Central Valley Water Board Executive Officer may revise this MRP Order as necessary, and the CRC shall comply with the MRP Order as revised by the Executive Officer.

The CRC, on behalf of the individual member Dischargers, shall implement the above monitoring and reporting program as of the date of this Order.

(original signed by)  
PAMELA C. CREEDON  
Executive Officer

02/17/09  
Date

- Order Attachment A** – Information Sheet
- Order Attachment B** – Definitions and Acronyms
- Order Attachment C** – Requirements for a Quality Assurance Project Plan
- Order Attachment D** – CRC MRP Plan – monitoring site descriptions and GPS coordinates

CALIFORNIA REGIONAL WATER QUALITY CONTROL BOARD  
CENTRAL VALLEY REGION

INFORMATION SHEET  
FOR  
ORDER NO. R5-2009-0809

MONITORING AND REPORTING PROGRAM  
FOR  
CALIFORNIA RICE COMMISSION  
UNDER AMENDED ORDER NO. R5-2006-0053  
COALITION GROUP CONDITIONAL WAIVER OF  
WASTE DISCHARGE REQUIREMENTS  
FOR DISCHARGES FROM IRRIGATED LANDS

## **I. REGULATORY BACKGROUND**

The California Regional Water Quality Control Board, Central Valley Region (Central Valley Water Board) adopts this Monitoring and Reporting Program (MRP) pursuant to California Water Code (Water Code) sections 13267 and 13269.

This Order is developed to conform to the “*Policy for Implementation and Enforcement of the Nonpoint Source Pollution Control Program*,” May 2004 (NPS Policy). The NPS Policy identifies five key elements that must be utilized by NPS implementation program as follows:

ELEMENT 1: An NPS control implementation program’s ultimate purpose shall be explicitly stated. Implementation programs must, at a minimum, address NPS pollution in a manner that achieves and maintains water quality objectives and beneficial uses, including any applicable anti-degradation requirements.

ELEMENT 2: An NPS control implementation program shall include a description of the MPs and other program elements that are expected to be implemented to ensure attainment of the implementation program’s stated purpose(s), the process to be used to select or develop MPs, and the process to be used to ensure and verify proper MP implementation.

ELEMENT 3: Where a RWQCB determines it is necessary to allow time to achieve water quality requirements, the NPS control implementation program shall include a specific time schedule, and corresponding quantifiable milestones designed to measure progress toward reaching the specified requirements.

ELEMENT 4: An NPS control implementation program shall include sufficient feedback mechanisms so that the RWQCB, dischargers, and the public can determine whether the program is achieving its stated purposes(s) or whether

additional or different MPs or other actions are required.

ELEMENT 5: Each RWQCB shall make clear, in advance, the potential consequences for failure to achieve an NPS control implementation program's stated purposes.

## II. MRP PLAN OBJECTIVES

The Irrigated Lands Regulatory Program (ILRP) oversees implementation of the terms and conditions of the Coalition Group Conditional Waiver, including development of the California Rice Commission (CRC) Monitoring and Reporting Program. Attachment B, Section B, Item 4 of the Conditional Waiver lists the objectives (purposes) of the CRC MRP. These objectives are consistent with the NPS Policy and include the following:

1. To determine whether the discharge of waste from irrigated lands within the Coalition Group boundaries causes or contributes to exceedances of applicable water quality standards or causes nuisance;
2. To provide information about the Coalition Group area characteristics, including but not limited to, land use, crops grown, and chemicals used;
3. To monitor the effectiveness of management practices implemented to address exceedances of applicable water quality standards;
4. To determine which management practices are most effective in reducing wastes discharged to surface waters from irrigated lands;
5. To specify details about monitoring periods, parameters, protocols, and quality assurance;
6. To support the development and implementation of the Conditional Waiver;
7. To verify the adequacy and effectiveness of the Conditional Waiver's conditions; and
8. To evaluate the Coalition Group's compliance with the terms and conditions of the Conditional Waiver.

The five Program questions listed below will assist the CRC in producing information to achieve these objectives. The CRC MRP and its associated Quality Assurance Project Plan (QAPP) and Management Plans shall be designed to address the five Program questions and ILRP objectives. The Central Valley Water Board recognizes that CRC Coalition has a long history in developing water quality programs, and that these five program questions will be useful for changes that have occurred over time, such as the use of newer products and other changes in management practices.

QUESTION No.1: Are conditions in waters of the State that receive discharges of wastes from irrigated lands within Coalition Group boundaries, as a result of activities within those boundaries, protective of beneficial uses?

QUESTION No.2: What is the magnitude and extent of water quality problems in waters of the State that receive agricultural drainage or are affected by other irrigated agriculture activities within Coalition Group boundaries, as determined using monitoring information?

QUESTION No.3: What are the contributing source(s) from irrigated agriculture to the water quality problems in waters of the State that receive agricultural drainage or are affected by other irrigated agriculture activities within Coalition Group boundaries?

QUESTION No.4: What are the management practices that are being implemented to reduce the impacts of irrigated agriculture on waters of the State within the Coalition Group boundaries and where are they being applied?

QUESTION No.5: Are water quality conditions in waters of the State within Coalition Group boundaries getting better or worse through implementation of management practices?

Implementation of this MRP Order will provide the information to determine whether rice field discharges are in compliance with the conditions of the Waiver, including compliance with applicable water quality standards.

### **III. REGULATORY BACKGROUND OF THE CALIFORNIA RICE COMMISSION**

The CRC and its predecessor, the California Rice Industry Association, have been working with the Central Valley Water Board, the State Regional Water Quality Control Board, the California Department of Fish and Game, and the Department of Pesticide Regulation since the early 1980's to establish practices for use of five rice pesticides to protect beneficial uses for waters of the State. As part of that work, the CRC worked with growers, registrants, and pest control advisors to establish practices such as hold times, application methods, and pesticide formulations for use of thiobencarb, molinate, carbofuran, malathion, and methyl parathion. Later the Central Valley Water Board approved these practices, and water quality goals were adopted in the Sacramento San Joaquin River Basin Plan. Monitoring for these pesticides continues under the Rice Pesticide Program (RPP), although only thiobencarb and molinate are still used on rice.

With the development of the Irrigated Lands Conditional Waiver, the CRC began to serve as a Coalition Group representing rice growers in the Sacramento Valley under the Coalition Group Conditional Waiver (Resolution R5-2003-015). The Coalition Group MRP Order R5-2003-0826 required the submittal of an MRP Plan by 1 April 2004. In 2004 the CRC worked with the Regional Board staff to develop a rice-specific MRP Order (CRC MRP Order No. R5-2004-0839, approved 18 November 2004) The CRC MRP Order has been revised over time, with an effort to retain some level of consistency with the Coalition Group Orders, with the exception of factors related to the commodity-specific nature of the CRC and its monitoring history.

This CRC MRP Order R5-2009-0809 is the most recent version, which is being revised in order to conform, where appropriate, to the Coalition Group MRP Order R5-2008-0005 (Coalition MRP Order), adopted in January 2008. This Information Sheet is intended to provide documentation of the technical rationale for differences between the Coalition MRP Order, and this CRC MRP Order.

The Coalition MRP Order allows flexibility in monitoring design provided that the objectives described in the Coalition MRP Order and in this Attachment A are met and that technical rationale is provided for any modifications to the monitoring design. This CRC MRP Order reflects an approach that will help address the MRP Plan objectives and meets the monitoring and reporting requirements in the Coalition Group MRP Order.

#### **A. CRC Monitoring History**

The design of the CRC MRP Order is specific to rice field discharges to waters of the State and includes a Monitoring Strategy that is consistent with the Coalition MRP Order in that it includes Assessment monitoring, Core monitoring, and Special Project monitoring components. The design also takes into account the information that has been gathered by the CRC since they began monitoring for water quality under the Irrigated Lands Regulatory Program in 2004, as well as information from previous activities conducted under the RPP.

The RPP monitoring originally sampled several sites in the Sacramento Valley watersheds. Eventually monitoring at four main RPP sites – Colusa Basin Drain above Knights Landing (CBD1), Colusa Basin Drain #5 (CBD5), Butte Slough at Lower Pass Road (BS1), and Sacramento Slough Bridge near Karnak (SSB) – were found to be most representative of rice field discharges for the five RPP pesticides of interest.

The CRC has been monitoring water quality and reporting monitoring results to the Central Valley Water Board in accordance with the Irrigated Lands Regulatory Program since 2004, utilizing the same four main monitoring sites from the RPP, but adding additional parameters required under the ILRP. Two additional sites, Jack Slough at Jack Slough Road (JS) and Lower Coon Creek at Striplin Road (LCC), were added to represent smaller water bodies and were monitored on a rotating basis for alternate years.

Table 1 below lists the monitoring sites that were sampled for the CRC from 2004 through 2008, as well as the site GPS coordinates.

<b>TABLE 1. CRC Monitoring Sites, 2004 – 2008</b>			
<b>Code</b>	<b>Site Name</b>	<b>GPS Coordinates</b>	
		<b>Latitude</b>	<b>Longitude</b>
CBD1	Colusa Basin Drain above Knights Landing	38.8125 N	-121.7731 W
CBD1	Colusa Basin Drain #5	39.1833 N	-122.0500 W
BS1	Butte Slough at Lower Pass Rd	39.1875 N	-121.9000 W
SSB	Sacramento Slough Bridge near Karnak	38.7850 N	-121.6533 W
JS	Jack Slough at Jack Slough Road	89.1804 N	-121.5711 W
LCC	Lower Coon Creek at Striplin Road	38.8715 N	-121.5808 W

The constituents analyzed included general parameters, toxicity, and at least two pesticides per year that were used only on rice fields. Water column sampling occurred monthly during the irrigation season, (April through September) and two water column samples were taken during the 'non-irrigation' season (February and October). The 'non-irrigation' season was scheduled to represent the storm season monitoring that was required by the Coalition MRP Order, and incorporated the winter drainage from rice fields. Winter drainage of rice fields is done to prepare for the winter flooding which begins winter straw decomposition.

Soil samples were also collected in July and September through 2005-07, and tested for sediment toxicity. Table 2 lists the parameters that were monitored by CRC from 2004 through 2008.

<b>TABLE 2. CRC Monitoring Parameters, 2004 – 2008</b>					
<b>Parameter</b>	<b>2004<sup>3</sup></b>	<b>2005</b>	<b>2006</b>	<b>2007</b>	<b>2008</b>
General Parameters <sup>1</sup>	√	√	√	√	√
Aquatic toxicity					
<i>Pimephales promelas</i>	√	√	√	√	√
<i>Ceriodaphnia dubia</i>	√	√	√	√	√
<i>Selenastrum capricornutum</i>	√	√	√	√	√
Sediment toxicity					
<i>Hyalella azteca</i>		√	√	√	
Pesticides					√
Azoxystrobin				√	Footnote 4
Bispyribac-sodium			√		Footnote 4
Carfentrazone ethyl			√		Footnote 4
Cyhalofop-butyl				√	Footnote 4
S-Cypermethrin	√	√			
Lambda Cyhalothrin	√	√			
Trifloxystrobin/Propiconazole				√	Footnote 4
Metals <sup>2</sup>			√		Footnote 4

<sup>1</sup> Includes pH, electrical conductivity, dissolved oxygen, temperature, color, turbidity, and flow.

<sup>2</sup> Includes for arsenic, boron, cadmium, copper, lead, nickel, selenium, zinc and hardness.

<sup>3</sup> Sampling in 2004 included only two events: September and October.

<sup>4</sup> Sampled under the Algae Toxicity Management Plan.

## **B. CRC Monitoring Design**

The CRC has developed a monitoring design that incorporates assessment monitoring, core monitoring and special project monitoring where needed. These are defined for the CRC MRP Order as follows:

**CRC ASSESSMENT MONITORING.** The assessment monitoring sites were chosen using a matrix system that evaluated individual subwatersheds, the total size of the drainage, and the acres of rice farmed in the subwatershed/drainage. The assessment monitoring sites were selected to achieve the following:

- provide data on water bodies representing a range of hydrologic conditions
- provide data to develop correlations between assessment and core sites
- confirm core site selection
- provide upstream data on new generation pesticides
- monitor water quality from drainages with a high percentage of land farmed in rice

**CRC CORE MONITORING.** The core monitoring sites were selected to track trends over extended periods of time. The four core sites have been extensively monitored for rice pesticides since the 1980's and were considered representative sites for the RPP pesticides. To determine if the core sites are still representative of rice discharges, monitoring data from assessment and core sites will be correlated. Every third year, the core sites will undergo assessment monitoring to evaluate the effects of changes in land-use and management practices and provide information about long-term trends and effectiveness of the management practices.

**CRC SPECIAL PROJECT MONITORING.** Special Project Monitoring includes monitoring for implementation of a TMDL and will also provide the mechanism for the CRC to implement Management Plans under Amended Order No. R5-2006-0053. A Management Plan is required when more than one exceedance of a parameter occurs at a site within a three-year period. The results from monitoring conducted under the CRC MRP Order indicate that water quality is impacted at some monitoring sites, and the Executive Officer has requested the preparation of a Management Plan to address these impacts. The Management Plan sites are identified as Special Project monitoring sites as described in this CRC MRP Order. The Special Project monitoring requirements are under development at this time.

## **C. CRC MONITORING PARAMETERS FOR 2009**

A narrative summary of the monitoring results from 2004-2008, and the use of these results to modify monitoring parameters and frequency that differ from the Coalition MRP Order are as follows:

**WATER COLUMN TOXICITY.** Water flea and fathead minnow testing did not result in exceedances sufficient to require a Management Plan, and monitoring for toxicity to these species will continue with the schedule for assessment monitoring. Algae toxicity

was found at each of the CRC monitoring sites and a Management Plan was triggered. Several unsuccessful attempts to determine the source of algae toxicity with Toxicity Identification Evaluations (TIEs) led to an alternative monitoring strategy that was proposed by the CRC in an Algae Toxicity Management Plan. The conditionally approved Algae Toxicity Management Plan proposed monitoring a suite of rice-specific herbicides, total copper and hardness with the schedule and site-locations based on exceedance patterns from previous years. The Management Plan also proposed to monitor four non-rice pesticides (glyphosate, simazine, diuron, and atrazine). The Management Plan is only conditionally approved at this time, and is undergoing revision by the Central Valley Water Board staff and the CRC.

SEDIMENT TOXICITY. No sediment toxicity has been found in four years of sampling. Samples were collected during July when rice fields may be releasing after peak pesticide use, and during the fall when fields are drained for harvesting. Monitoring for sediment toxicity under this CRC MRP Order will take place one time per year (late September) at assessment sites, after the completion of fall drainage which is the most likely season to capture sediment toxicity.

METALS. Two metals are applied to rice fields; copper used as a fungicide and zinc as a micronutrient. In 2006, samples were analyzed for arsenic, boron, copper, hardness, lead, nickel, selenium, and zinc. Metals analyzed were not detected above the water quality criteria or objective. In 2007, copper and hardness were analyzed in July and August as described in the conditionally-approved Management Plan for Algae Toxicity.

GENERAL PARAMETERS AND NUTRIENTS. This category in the Coalition MRP Order includes field parameters such as temperature, dissolved oxygen (DO), pH, electrical conductivity (EC), TOC, *E. coli*, turbidity and nutrients such as ammonia, nitrates, and phosphorous.

Several general parameters such as total dissolved solids (TDS), EC, total organic carbon (TOC) and dissolved organic carbon (DOC), are part of the UC Davis study that uses edge-of-field monitoring. Nutrients such as nitrogen and phosphorus are generally incorporated into the seedbed before flooding, although both may be applied later to correct deficiencies and maintain plant growth and yield. Nutrient monitoring was conducted as part of the UC Davis edge-of-field study.

The CRC proposes to use the results of the UC Davis study to help determine if nutrients, TDS, water column TOC and other parameters will need to be monitored as part of the Core and Assessment suite of analytes in the future. Available results from the UC Davis study will be presented and discussed in the CRC 2008 Annual Monitoring Report. The final study from UC Davis is due, by contract, on 31 March 2009, and will be evaluated by the Central Valley Water Board to determine if monitoring should be added to a revised CRC MRP Plan.

Animal waste, such as manure or wastewater, is never applied to rice fields. Therefore, monitoring for pathogens is not a requirement of this CRC MRP Order. However, if the Sacramento Valley Water Quality Coalition is developing a pathogen management plan for a water way receiving rice drainage, the CRC may be requested to participate in any source identification studies.

PHOTO MONITORING. The monitoring sites identified in the CRC MRP Order are well documented sites that seldom change in appearance, with some seasonal differences caused by storm events. Photo documentation will occur at all sites initially to identify "normal" conditions, and as needed to document site changes that could affect monitoring results or the ability to monitor (such as dry conditions). Digital cameras will be part of the sampling crew equipment so that site changes can be documented where necessary, but they will only be used when necessary.

PESTICIDE MONITORING. From 2004-2007, the CRC monitored at least two rice-specific pesticides every year. To select the pesticides, the CRC performed an extensive evaluation in consultation with Regional Board staff of the pesticides that involved review of the acreage applied; physical characteristics such as solubility, half-life in soil and water, and soil/water coefficient; and whether the pesticide had been detected at levels of concern in any previous sampling.

For this CRC MRP Order, the CRC performed the same evaluation on currently used pesticides, using more up-to-date information on usage, aquatic toxicity, and detection in surface water and past monitoring locations. To determine which rice-specific pesticides would be incorporated into this CRC MRP Order, the information provided on Tables 3 and 4 were used. DPR data for rice pesticide use in 2007 is shown in Table 3. Each pesticide is ranked according to pounds active ingredient applied (Rank 1), number of acres applied (Rank 2), and number of applications for the year (Rank 3) based on all pesticides and applications performed in 2007. The pesticides listed in Table 3 comprise 83% of all active ingredients (active component of the pesticide) applied to rice, and 70% of all pesticide applications. Pesticides with less than 1000 pounds of active ingredient applied in 2007 were not examined. Copper will be analyzed as part of the metal analyses. When copper is excluded, the fifteen listed pesticides will cover almost 75% of all active ingredients applied to rice, and 68% of all pesticide applications.

Table 4 uses the list of pesticides in the previous table and provides the technical rationale for the monitoring decision. Selection for monitoring was based on the same criteria (acreage, pounds applied, solubility, half-life, LC or EC 50s, etc) using pesticide use data from 2007 and the most recent data on aquatic toxicity from the USEPA ECOTOX database. The ECOTOX database can be found at the following link: <http://cfpub.epa.gov/ecotox>.

In Table 4, the sixteen pesticides, including copper, were examined based on pounds of active ingredient divided by the acres applied. This ranking can be used as an indicator

of the probability to detect any releases from rice fields. Although other pesticides may have higher ratios, the total number of pounds applied (less than 1000 pounds for the year) and the low acreage would likely make detection highly improbable at the selected monitoring sites.

The exception to this technical approach is made for propanil, for the interim. Based on the pounds of active ingredient, acreage applied, and ECOTOX data, propanil should be analyzed. However, the CRC is aware of data prepared by the registrant of propanil, combined with CRC hold times that may indicate that this product does not have the potential to impact water quality. The CRC will submit their registrant data to the Central Valley Water Board in the December 2008 Annual Monitoring Report with a discussion regarding the need to monitor propanil. At that time the CRC will coordinate a meeting with the registrants and the Regional Water Board staff to determine the next steps and whether or not to add propanil to the Assessment Monitoring requirements for the CRC MRP Order.

#### **D. SPECIAL PROJECT MONITORING**

The requirement for a plan to address *Selenastrum capricornutum* (algae) toxicity was noted by the CRC in its 2006 Annual Monitoring Report (AMR). On 27 August 2007, the CRC was sent a revised table and summary on CRC water quality exceedances. During the year, the CRC and Central Valley Water Board staff discussed a strategy that could address the *Selenastrum capricornutum* toxicity at CRC monitoring sites. The Executive Officer (EO) gave conditional approval to the Algae Toxicity Management Plan on 9 June 2008. Conditions of the approval included submittal of method validation for non-standard procedures used in testing of specific pesticides listed in the proposed Management Plan. This information has not yet been submitted, and the CRC will be proposing an alternative Management Plan strategy.

#### **E. REPORTING**

The CRC will be submitting one annual report in December, rather than in the month of March, which is the submittal date in the Coalition MRP Order. This is being done in order to maintain consistency with the CRC reporting schedule for the RPP.

**TABLE 3. Pesticides used by Rice Production -- 2007 DPR data**

Name	Pounds of Active Ingredient (AI)	Rank1	Acres Applied	Rank2	Number of Applications	Rank3	Pounds AI/acres applied	Type of Pesticide
Propanil	1,801,285	1	377,903	1	5,173	1	4.8	Herbicide
Copper (all forms)	1,647,272	2	127,024	5	1,784	4	13.0	Algaecide/ Fungicide/ Insecticide
Thiobencarb	289,033	3	74,251	8	804	11	3.9	Herbicide
Clomazone	79,711	4	159,161	3	2,043	3	0.5	Herbicide
Molinate	75,235	5	17,471	13	212	15	4.3	Herbicide
Triclopyr TEA	49,319	6	295,644	2	3,844	2	0.2	Herbicide
Cyhalofop butyl	36,527	7	119,979	6	1,687	5	0.3	Herbicide
Azoxystrobin	22,170	8	139,787	4	1,634	6	0.2	Fungicide
Pendimethalin	8,644	9	9,946	15	119	17	0.9	Herbicide
2,4-D dimethylamine salt	5,218	10	19,946	12	262	13	0.3	Herbicide
Glyphosate (all forms)	5,157	11	6,135	16	98	18	0.8	Herbicide
Penoxsulam	2,962	12	82,492	7	1,064	9	0.036	Herbicide
(s) Cypermethrin	2,310	13	48,412	11	1,219	8	0.048	Insecticide
Carfentrazone ethyl	2,007	14	16,090	14	223	14	0.125	Herbicide
Bispyribac sodium	1,730	15	62,019	9	1,018	10	0.028	Herbicide
Lambda-Cyhalothrin	1,679	16	59,505	10	1,370	7	0.028	Insecticide
Other rice pesticides*	< 1000							

\*Other pesticides used on rice are at lower levels of application; less than 1000 pounds each per year

	Total for above	Total for all rice	% pesticides in table compared to all rice pesticides	Total for above excluding copper	Total for all rice excluding copper	% pesticides compared to all rice pesticides, excluding copper
Pounds of Active Ingredient	4,026,704	4,839,890	83%	2,382,987	3,196,173	74.5%
Number of Applications	22,546	32,337	70%	20,770	30,561	68%

**TABLE 4. Pesticides Examined and Rationale for Monitoring Decisions: 2009**

Monitor?		Chemical Name	Pounds Active Ingredient per acre applied	Rationale
YES	NO			
√		Copper sulfate	13.0	Will be monitored as part of metal scan at assessment sites.
	√	Propanil	4.8	Registrant information to be reviewed in 2009 and monitoring to be considered for future years.
	√	Molinate	4.3	Monitored is conducted and reported under the RPP.
	√	Thiobencarb	3.9	Monitored is conducted and reported under the RPP.
√		Pendimethalin	0.9	To be monitored at core and assessment sites. No monitoring data available.
√		Glyphosate	0.6	Used for border treatment to control weeds surrounding rice fields. To be monitored at core and assessment sites.
√		Clomazone	0.5	New general pesticide showing increasing use on rice. Will be used to determine if there is sufficient correlation between detections at core and assessment sites. 14 day hold time
	√	Cyhalofop-butyl <sup>1</sup>	0.3	Short half-life when pH > 7; 7 day hold time. High adsorption to soils. May be evaluated if toxicity tests indicate that it is warranted.
	√	2,4-D dimethylamine salt	0.3	High aquatic toxicity values in the ppm range, which are not likely to result from rice applications. High volumes of application are used by non-agricultural sources.
	√	Azoxystrobin	0.2	14 day hold time. Generally applied in early stages of rice development and not released. High volumes of application are used by non-rice sources. To be evaluated if toxicity tests indicate that it is warranted.
	√	Triclopyr <sup>2</sup>	0.2	High aquatic toxicity values in the ppm range. High volumes of application by non-rice and non-agricultural sources.
√		Carfentrazone-ethyl	0.125	Will be used to determine if there is sufficient correlation between detections at core and assessment sites.
√		S-Cypermethrin	0.048	Pyrethroid; binds to soil and is generally not found in water column. This pesticide will be analyzed if sediment toxicity is found.
√		Penoxsulam	0.036	To be monitored. No monitoring data available.
	√	Bispyribac-sodium <sup>3</sup>	0.028	CRC is required to hold the pesticide on the field for 7 days before release of water; low rate of application (1,730 pounds for 62,019 acres)
√		Lambda-Cyhalothrin	0.028	Pyrethroid, binds to soil and generally not found in water column. This pesticide will be analyzed if sediment toxicity is found.

<sup>1</sup> EC 50 of 580 ppb for rainbow trout.

<sup>2</sup> EC 50 of 5900 ppb for green algae. Triclopyr TEA immediately converts to triclopyr (same as triclopyr acid) in water.

<sup>3</sup> EC 50 of 2760 ppb for green algae.



## ATTACHMENT B

### CALIFORNIA REGIONAL WATER QUALITY CONTROL BOARD CENTRAL VALLEY REGION

#### MONITORING AND REPORTING PROGRAM ORDER NO. R5-2009-00809

#### APPLICABLE DEFINITIONS AND ACRONYMS FOR CALIFORNIA RICE COMMISSION FOR DISCHARGES FROM IRRIGATED LANDS

The following information is presented to provide definition and clarification of terminology and acronyms used within the Monitoring and Reporting Program documents.

#### **Definitions**

The following definitions apply to the Monitoring and Reporting Program as related to discharges from irrigated lands as described in this Order and all attached documents.

1. **Accuracy** - The closeness or agreement of the observed value or test response to the true or acceptable reference value or the test response from a reference method. It is influenced by both random error (precision) and systematic error (bias). The terms "bias" and "precision" are often used in lieu of "accuracy".
2. **Analytical Batch** - A group of 20 or fewer samples and associated quality control that is processed by the same instrument within a 24-hour period. Multiple sample batches can comprise an analytical batch.
3. **Analytical Run** - The quantification of a single discrete sample or its associated quality control.
4. **Assessment** - A general evaluation process used to evaluate the performance, effectiveness, and processes of a management and/or technical system.
5. **Batch** - A group of 20 or fewer samples, to include quality control samples, which is to be collected and/or analyzed in one, test run or inspected together within a specific time limit and traceable as a unit.
6. **Bias** - The constant or systematic distortion of a measurement process that manifests itself as a persistent positive or negative deviation from the known or true value. This can result from improper data collection, poorly calibrated analytical or sampling equipment, or limitations or errors in analytical methods and techniques.
7. **Blank** - A specimen that is intended to contain none of the analytes of interest and which is subjected to the usual analytical or measurement process to establish method purity, a zero baseline, or background value.

- 8. Calibration** - A comparison of a measurement standard, instrument, or item with one having higher accuracy to detect, quantify, and record any inaccuracy or variation; the process by which an instrument setting is adjusted based on response to a standard to eliminate the inaccuracy.
- 9. Calibration Standard** - A reference solution or substance of known value or chemical concentration used to establish a correct instrument reading.
- 10. Certified Reference Materials** - A substance or solution for which the composition or concentration of a particular chemical constituent is known, and which is traceable with documentation pertaining to its composition and uniformity to an established standardization organization such as the National Institute for Standards and Technology (NIST) or the American Association for Laboratory Accreditation (A2LA).
- 11. Chain-of-Custody** - An unbroken, documented trail of accountability that ensures the physical security and/or integrity of samples, data, and records.
- 12. Coalition Group** - A group of dischargers and/or organizations that choose to comply with the Conditional Waiver by forming a group which is approved by the Central Valley Regional Water Quality Control Board. Coalition Groups can be organized on a geographic basis or can be groups with other factors in common such as commodity groups.
- 13. Coefficient of Variation** - The standard deviation divided by the mean; a unit-free measure of variability.
- 14. Comparability** - A measure of the confidence with which one data set, element, or method can be considered as similar to another, e.g., taken from the same location, taken in a similar manner, etc.
- 15. Completeness** - A measure of the amount of valid data obtained from a measurement system, compared to the planned or expected amount. For the ILRP, completeness goals will be evaluated with the submittal of each annual monitoring report. The completeness evaluation will include the number of samples successfully obtained and the proportion of quality control samples that are within acceptance criteria.
- 16. Contamination** - The unintentional addition of analytical constituents to a sample or system.
- 17. Continuing Calibration Verification** - A periodic standard used to assess instrument drift between calibrations.
- 18. Control Chart** - A graphic representation of the variability in a measurement process generally plotted in order over time.
- 19. Control Limit** - The upper and lower acceptable ranges of process data used to judge whether the process is within or outside of statistical limitations. Control limits are determined by the variation in a process data set expressed as the

mean value plus or minus a pre-determined number of standard deviations (typically three standard deviations from the mean).

- 20. Corrective Action** - Any measures taken to rectify conditions adverse to quality and/or to eliminate the causes of an existing nonconformity, defect, or other undesirable situation in order to prevent reoccurrence.
- 21. Data Quality Assessment** - A statistical and scientific evaluation of a data set to determine the validity and performance of the data collection design and execution, and to determine the adequacy of the data set for its intended use.
- 22. Data Quality Indicators** - The quantitative statistics and qualitative descriptors that are used to interpret the degree of acceptability or utility of information to the user. The principal DQIs are precision, accuracy (or bias), representativeness, comparability, completeness, and sensitivity.
- 23. Data Quality Objectives** - Qualitative and quantitative statements derived from the DQO Planning Process that clarify the purpose of the study, define the most appropriate type of information to collect, determine the most appropriate conditions from which to collect that information, and specify tolerable levels of potential decision errors.
- 24. Data Quality Objectives Process** - A systematic strategic development tool based on the scientific method that identifies and defines the type, quality, and quantity of information needed to satisfy a specified use, including data precision, accuracy, and completeness requirements.
- 25. Data Validation** - An analyte- and sample-specific process that evaluates analytical information after the verification process (i.e., determination of method, procedural, or contractual compliance) to determine analytical quality and any limitations on the data.
- 26. Data Verification** - The process of evaluating the completeness, correctness, and conformance/compliance of a specific information set against the method, procedural, or contractual specifications for that activity.
- 27. Discharger** - The owner and/or operator of irrigated lands or a Water District, which accepts or receives discharges from irrigated lands, who discharges or threatens to discharge: irrigation return flows, tailwater, operational spills, drainage water, subsurface drainage generated by irrigating crop land or by installing drainage systems to lower the water table below irrigated lands (tile drains) and/or stormwater runoff flowing from irrigated lands to waters of the State.
- 28. Discharges from irrigated lands** - Include surface discharges (also known as irrigation return flows or tailwater), operational spills, drainage water discharges, subsurface discharges through drainage systems that lower the water table below irrigated lands (also known as tile drains), stormwater runoff flowing from irrigated lands, and stormwater runoff conveyed in channels or canals resulting from the discharge from irrigated lands. For the purpose of this Coalition Group

Monitoring and Reporting Program, stormwater discharges to surface waters resulting from any size storm can be covered by this Conditional Waiver.

- 29. Drift** - The deviation in instrument response from its set or reference value over a period of time.
- 30. Equipment Blank** - An aliquot of reagent water that is subjected to all aspects of sample collection and analysis, including contact with all sampling devices and apparatus. The purpose of the equipment blank is to determine if the sampling devices and apparatus for sample collection have been adequately cleaned prior to use.
- 31. Field Blank** - An aliquot of reagent water which is exposed to sampling conditions, returned to the laboratory, and treated as an environmental sample. This blank is used to provide information about contaminants that may be introduced during sample collection, storage, and transport.
- 32. Field Duplicate (Co-located)** - An independent specimen collected from (as closely as possible) the same point in time and space as the primary specimen. This would include duplicate sample containers filled simultaneously and in close proximity to one another from the same medium, or duplicate containers filled in rapid succession from the same location or source.
- 33. Field Duplicate (Sub-sample) or Field Split** - A test specimen that is homogenized before being divided into two or more portions with the same laboratory analyzing all portions, to evaluate sampling and analysis precision. This type of field duplicate (or split) sample analysis can also be performed by more than one lab to evaluate inter-laboratory precision.
- 34. Field Measurements** - Those activities associated with performing analyses or measurements in the habitat being examined.
- 35. Holding Time** - The period of time a sample may be stored following collection, preservation, extraction, or analysis. While exceeding the holding time does not necessarily negate the validity of analytical results, associated analytical data are typically qualified as estimated.
- 36. Indicators** - Items, elements, or measures used to determine or identify a basic condition or how well a process or program is meeting its objectives.
- 37. Inter-comparison** - An exercise in which samples are prepared and split by a reference laboratory, then analyzed by one or more testing laboratories and the reference laboratory. The inter-comparison, with a reputable laboratory as the reference laboratory, serves as a test of the precision and accuracy of the analyses from different laboratories at natural environmental levels.
- 38. Interference** - An element, compound, or other matrix effect present in a sample, which disturbs the detection of a target analyte leading to inaccurate concentration results for the target analyte.

- 39. Internal Standard** - Pure analyte (s) added to a sample, extract, or standard solution in known amount(s) and used to measure the relative responses of other method analytes that are components of the same sample or solution. The internal standard must be an analyte that is not a sample component.
- 40. Irrigated Lands** - Lands where water is applied for the purpose of producing crops, including, but not limited to, land planted to row, vineyard, pasture, field and tree crops, commercial nurseries, nursery stock production, managed wetlands, rice production, and greenhouse operations with permeable floors that do not currently discharge under waste discharge requirements (WDRs), including Municipal Separate Storm Sewer System or other National Pollutant Discharge Elimination System permits are considered irrigated lands.
- 41. Irrigation Season** - The time of year when water is applied to fields for the purpose of promoting crop growth, for distributing nutrients or other chemicals to crop lands or for the purposes of counteracting the effects of frost during cold season months.
- 42. Irrigation Return Flow** - Surface and subsurface water that leaves the field following application of irrigation water.
- 43. Laboratory Blank (also known as a Method Blank)** - An aliquot of reagent water (or for solid matrices, an inert solid similar to the sample matrix) that is prepared by the laboratory and treated exactly as a sample, including exposure to all glassware, equipment, solvents, reagents, internal standards, and surrogates that are used with samples. The laboratory blank is used to determine if method analytes or interferences are present in the laboratory environment, the reagents, or the apparatus.
- 44. Laboratory Duplicate** - Two or more representative portions taken from one homogeneous sample by the laboratory analyst and analyzed in the same testing facility to evaluate the effects of laboratory conditions on analytical precision.
- 45. Laboratory Control Sample** - A specimen of known composition prepared using contaminant-free reagent water, or an inert solid, that is spiked with the analyte of interest at the midpoint of the calibration curve or at the level of concern; and then analyzed using the same preparation, reagents, and analytical methods employed for regular specimens and at the intervals set in the Quality Assurance Project Plan.
- 46. Matrix** - The material of which the sample is composed or the substrate containing the analyte of interest, such as drinking water, waste water, air, soil/sediment, biological material, etc. Also called medium or media.
- 47. Matrix Spike** - A test specimen that is prepared by adding a known concentration of the target analyte(s) to a specified amount of a specific homogenized specimen and is then subjected to the entire analytical protocol.

- 48. Matrix Spike Duplicate** - A sample prepared simultaneously as a split with the matrix spike sample with each specimen being spiked with identical, known concentrations of targeted analyte.
- 49. Measurement Quality Objectives** - The individual performance or acceptance goals (or requirements) for the individual Data Quality Indicators such as precision or bias.
- 50. Metadata** - The information about a data set, which may include descriptive information about the context, quality and condition, or characteristics of a data set. For geographical data this may include the source of the data; its creation date and format; its projection, scale, resolution, and accuracy; and its reliability with regard to some standard.
- 51. Method** - A procedure, technique, or tool for performing a scientific activity.
- 52. Method Detection Limit** - The minimum concentration of an analyte that undergoes the entire measurement process and can be reported with a stated level of confidence that the analyte concentration is greater than zero.
- 53. Method Linearity** – The ability of an analytical method to demonstrate an increase in sample concentration of a given analyte, as the instrument response also increases. Demonstration of instrument linearity, as well as the upper and lower limits of linearity, are considered part of a laboratory method validation procedure and should take place before the procedure is used to report analytical results.
- 54. Monitoring** - All types of monitoring undertaken in connection with determining water quality conditions and factors that may affect water quality conditions, including but not limited to, in-stream water quality monitoring undertaken in connection with agricultural activities, monitoring to identify short and long-term trends in water quality, active inspections of operations, and management practice implementation and effectiveness monitoring.
- 55. Negative Control** - Measures taken to insure that a test, its components, or the environment do not cause undesired effects, or produce incorrect test results.
- 56. Operational Spill** – Irrigation water that is diverted from a source such as a river, but is discharged without being delivered to or used on an individual field.
- 57. Parameter** - A statistical quantity, usually unknown, such as a mean or a standard deviation, which characterizes a population or defines a system. The term Parameter (or sometimes “Analytical Parameter”) can also be defined as a measured analytical constituent such as an individual chemical, a group of chemicals, or a physical property (i.e. Total Organic Carbon, electrical Conductivity, etc.).
- 58. Performance Based Measurement System** - A set of processes wherein the data needs, mandates, or limitations of a program or project are specified and

serve as criteria for selecting appropriate methods to meet those needs in a cost-effective manner.

- 59. Positive Control** - A prepared standard which undergoes an analytical procedure to provide comparison with an unknown specimen thereby monitoring recovery to assure that a test and/or its components are working properly and producing correct or expected results.
- 60. Precision** - A measure of mutual agreement between two or more individual measurements of the same property, obtained under similar conditions.
- 61. Proficiency Test** - A type of external assessment in which a stable sample, the composition of which is unknown to the analyst, is provided to determine whether the analyst/laboratory can produce analytical results within the specified acceptance criteria. Also known as a Performance Evaluation Test.
- 62. Proficiency Test Sample** - A test specimen of known composition and/or chemical concentration that mimics an actual specimen in all possible aspects, except that its composition is unknown to the laboratory at the time of analysis, and which is used to assess the laboratory's capability to produce results within acceptable criteria.
- 63. Qualified Data** - Any numerical information that may be of limited use for a specific function, and is identified (flagged) as such.
- 64. Quality Assurance** - An integrated system of management activities (planning, implementation, assessment, reporting, and quality improvement) that focuses on providing confidence in the data or product by ensuring that it is of the type and worth needed and expected for its expressed, intended use.
- 65. Quality Assurance Officer** - The individual designated within an organization having management oversight and responsibilities for planning, documenting, coordinating, and assessing the system effectiveness for ensuring the value of the work.
- 66. Quality Assurance Project Plan** - A document that describes the intended technical activities and project procedures that will be implemented to ensure that the results of the work to be performed will satisfy the stated performance or acceptance criteria. The amount of information presented and the planned activities to ensure the value of the work will vary according the type of study and the intended use of the data.
- 67. Quality Control** - The overall system of technical activities that measures the attributes and performance of a process, item, or service against defined standards to verify that they meet the stated requirements established; operational techniques and activities that are used to fulfill requirements.
- 68. Quality Control Sample** - One of any number of test specimens, such as a Proficiency Test or blank, intended to demonstrate that a measurement system or activity is in check.

- 69. Quality Management Plan** - A document that describes an organization's system in terms of its organizational structure, policy and procedures, staff functional responsibilities, lines of authority, and interfaces for those planning, implementing, documenting, and assessing all activities conducted.
- 70. Quality Objectives** - The combined characteristics of Data Quality Objectives and Measurement Quality Objectives; the overall criteria related to sample design and analytical measurements intended to assure that analytical data meet the requirements associated with the intended use.
- 71. Quantitation Limit or Practical Quantitation Limit (PQL)** - The level above which numerical results may be obtained with a specified degree of confidence, the minimum concentration of an analyte, or category of analytes, in a specific matrix that can be identified and quantified within specified limits of precision and accuracy during routine analytical operating conditions. The manner of establishing the quantitation limit is method-specific, and typically involves the successful (within established acceptance criteria) analysis of calibration standards at the quantitation limit concentration -- either as part of the instrument calibration procedure, or as a routine control sample.
- 72. QC Set (Quality Control Set)** - A group of quality control samples (i.e. a method blank, a matrix spike and matrix spike duplicate, etc.) used to evaluate (control) a specific set or batch of samples.
- 73. Receiving waters** - Surface waters that receive or have the potential to receive discharges from irrigated lands.
- 74. Recovery** - The measure of accuracy for an analytical procedure, including determining whether or not the methodology measures all of the analyte contained in a sample, often expressed in percent recovered.
- 75. Reference Toxicant** - A substance used as a positive control for toxicological analyses to test the sensitivity of the test organisms to a known toxic substance, and to assure appropriate lab procedures have been performed.
- 76. Relative Percent Difference** - The absolute value of the difference of two measurements divided by the statistical mean of the same two measurements, used to evaluate the precision of duplicate samples analysis, or two repeated measurements.
- 77. Relative Standard Deviation** - The standard deviation divided by the mean; a unit-free measure of variability.
- 78. Repeatability** - The degree of agreement between independent test results produced by the same analyst, using the same test method and equipment on random aliquots of the same sample within a short time period.
- 79. Reporting Limit (RL)** - the quantitation level required by the Irrigated Lands Program for reporting purposes. The RL is typically set at a laboratory quantitation level, but consideration may be made for lowering the level to the

detection limit, if information about presence or absence of a contaminant is necessary. Similarly, if levels that are protective of water quality prove to be lower than the routine quantitation limit at a given laboratory, then the CVRWQCB may require an RL that is lower than the PQL, providing achieving that limit is economically feasible. The RL can sometimes be raised to some default value above the PQL, if the PQL is much lower than necessary to protect water quality, and if it is approved by the CVRWQCB.

- 80. Representativeness** - A measure of the degree to which data accurately and precisely represent characteristics of a population, parameter variations at a sampling point, a process condition, or an environmental condition.
- 81. Rinse Blank** - A dilute acid solution used to flush an instrument between samples in order to reduce memory interferences.
- 82. Sample Batch** - A group of 20 samples or fewer and associated quality control that is collected by the same entity within a 24-hour period.
- 83. Sensitivity** - The capability of a method or instrument to discriminate between measurement responses representing different levels of a variable of interest.
- 84. Spike** - A known quantity of an analyte added to a sample for the purpose of determining recovery or efficiency (analyst spikes), or for quality control (blind spikes).
- 85. Split** - Two or more representative portions taken from one specimen in the field or in the laboratory and analyzed by different analysts, methods, or laboratories.
- 86. Standard Deviation** - The measure of the dispersion or imprecision of a series of accepted results around the average, equal to the square root of the variance.
- 87. Standard Operating Procedure** - A written document that details the method for an operation, analysis, or action with thoroughly prescribed techniques and steps and that is officially approved as the method for performing certain routine or repetitive tasks.
- 88. Stormwater runoff** – The runoff of precipitation from irrigated lands to surface waters from any size storm event.
- 89. Subsurface drainage** – Water generated by installing drainage systems to lower the water table below irrigated lands. Subsurface drainage systems, deep open drainage ditches, or drainage wells can generate this drainage.
- 90. Surrogate** - A pure substance with properties that mimics the analyte of interest (organics only) and which is unlikely to be found in environmental samples. It is added into a sample before sample preparation.
- 91. Tailwater** – The runoff of irrigation water from an irrigated field.
- 92. Travel Blank** - Analyte-free water placed in the same type of container as its associated field samples. It may be pre-preserved prior to shipment, but is not

opened during the sample collection. Consequently, it helps isolate contamination associated with sample transport.

- 93. Waste** – As defined in California Water Code (Water Code) Section 13050. Includes sewage and any and all other waste substances, liquid, solid, gaseous, or radioactive, associated with human habitation, or of human or animal origin, or from any producing, manufacturing, or processing operation, including waste placed within containers or whatever nature prior to, and for the purposes of disposal. Waste specifically regulated by the Coalition Group Conditional Waiver includes: earthen materials, such as soil, silt, sand, clay, and rock; inorganic materials, such as metals, salts, boron, selenium, potassium, nitrogen, etc.; and organic materials, such as pesticides that enter or threaten to enter waters of the State. Examples of waste not specifically regulated by the Coalition Group Conditional Waiver include hazardous and human wastes.
- 94. Water Quality Standards** – Water Quality Standards consist of narrative and numeric water quality objectives in the Central Valley Regional Water Quality Control Board's Basin Plans, water quality criteria in the California Toxics Rule and National Toxics Rule adopted by the USEPA, and/or water quality objectives in other applicable State Water Board plans and policies.
- 95. Waters of the State** – As defined in Water Code Section 13050. Any surface water or groundwater, including saline waters, within the boundaries of the State. This Order and the Coalition Group Conditional Waiver currently regulate only discharges from irrigated lands to surface waters.

## Acronyms

The following acronyms apply to the Monitoring and Reporting Program as related to discharges from irrigated lands as described in this Order and all attached documents.

AMR	Annual Monitoring Report
CAL-EPA	California Environmental Protection Agency
CCR	California Code of Regulations
CFR	Code of Federal Regulations
COC	Chain of Custody
CTR	California Toxics Rule
CWA	Clean Water Act
DFG	Department of Fish and Game
DHS	Department of Health Services
DO	Dissolved Oxygen
DOC	Dissolved Organic Carbon
DPR	Department of Pesticide Regulation
DQO	Data Quality Objective
DWR	Department of Water Resources
GC/MS	Gas chromatography/mass spectrometry
IDL	Instrument Detection Limit
GIS	Geographic Information System
ILRP	Irrigated Lands Regulatory Program
LCS	Laboratory Control Spike
LCSD	Laboratory Control Spike Duplicate
LTMS	Long-term Monitoring Strategy
ML	Minimum Level
MCL	Maximum Contaminant Level
MDL	Method Detection Limit
MRP	Monitoring and Reporting Program
MRPP	Monitoring and Reporting Program Plan
MP	Management Practices
MS	Matrix Spike
MSD	Matrix Spike Duplicate
MUN	Municipal use of a water body as a source of drinking water
N/A	Not Applicable
NPDES	National Pollutant Discharge Elimination System
NRCS	Natural Resources Conservation Service
NTR	National Toxics Rule
ppm	Parts per million (mg/kg sediment and tissue; mg/l water)
ppb	Parts per billion (ug/kg or ng/g sediment and tissue; ug/l water)
PQL	Practical Quantitation Limit
QAMP	Quality Assurance Management Plan
QAPP	Quality Assurance Project (or Program) Plan
QA/QC	Quality Assurance/Quality Control
QO	Quality Objective

REC1	Contract recreation as a beneficial use for a water body
RL	Reporting Limit
RPD	Relative Percent Difference
RWQCB	Regional Water Quality Control Board
SAMR	Semi-annual Monitoring Report
SD	Standard Deviation
SOP	Standard Operating Procedure
SWAMP	Surface Water Ambient Monitoring Program
SWRCB	State Water Resources Control Board
SVOC	Semi-volatile organic carbon compounds
TIE	Toxicity Identification Evaluation
TKN	Total Kjeldahl Nitrogen
TMDL	Total Maximum Daily Load
TOC	Total Organic Carbon
TRL	Target Reporting Limit
TSS	Total Suspended Solids
USEPA	United States Environmental Protection Agency
USGS	United States Geological Survey
WER	Watershed Evaluation Report
VOA	Volatile Organic Analysis
VOC	Volatile Organic Compounds

**CALIFORNIA REGIONAL WATER QUALITY CONTROL BOARD  
CENTRAL VALLEY REGION**

**QUALITY ASSURANCE PROJECT PLAN GUIDELINES  
FOR  
CALIFORNIA RICE COMMISSION**

**ORDER NO. R5-2009-0809  
MONITORING AND REPORTING PROGRAM  
UNDER AMENDED ORDER NO. R5-2006-0053  
COALITION GROUP CONDITIONAL WAIVER OF  
WASTE DISCHARGE REQUIREMENTS  
FOR DISCHARGES FROM IRRIGATED LANDS**

## IRRIGATED LANDS CONDITIONAL WAIVER PROGRAM QUALITY ASSURANCE PROJECT PLAN GUIDELINES

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## **IRRIGATED LANDS CONDITIONAL WAIVER PROGRAM QUALITY ASSURANCE PROJECT PLAN GUIDELINES**

### **I INTRODUCTION**

A Quality Assurance Project Plan (QAPP) shall be developed by the Discharger and shall include site-specific information and field and laboratory quality assurance requirements. This document identifies the major elements of the quality assurance and quality control (QA/QC) components that need to be described in the QAPP. The QAPP shall be submitted to the staff of the Central Valley Water Board Irrigated Lands Conditional Waiver Regulatory Program (ILRP) for review and approval by the Central Valley Water Board Quality Assurance Officer.

### **II OBJECTIVE**

The purpose of this document is to identify the QA and QC components that must be described in the QAPP for the Discharger monitoring. A QAPP contains the requirements and criteria for the field and laboratory procedures used during planning and implementation of the monitoring program. The QAPP shall identify the procedures that will be used to assure that the monitoring data represents, as closely as possible the water quality conditions of the water body that is being sampled at the time of sampling. This will be achieved by using accepted methodologies (e.g., U.S. Environmental Protection Agency, USEPA) for sample collection and analysis of water, sediment, and biota. Chemical, bacteriological, and bioassay analyses shall be conducted at a laboratory certified for such analyses by the State Department of Health Services. In the event a certified laboratory is not available to the discharger, analyses performed by a non-certified laboratory will be accepted provided a Quality-Assurance Quality Control Program is instituted by the laboratory. A manual containing the steps followed in this program must be kept in the laboratory and shall be available for inspection by Board staff. The Discharger's ability to meet this objective will be assessed by evaluating the monitoring detection limits, precision, accuracy, comparability, representativeness, and completeness. A QAPP must contain adequate detail for project and Water Board staff to identify and assess the technical and quality objectives, measurement and data acquisition methods, and limitations of the data generated under the project. This document provides a description of major elements of a QAPP that are also required under the guidelines provided by the USEPA and the State Surface Water Ambient Monitoring Program (SWAMP).

Note: This document provides a compilation of USEPA, SWAMP and ILRP guidelines. Language has been taken and used directly from the following documents:

USEPA. 2001 (2006) USEPA Requirements for Quality Assurance Project Plans (QA/R-5) Office of Environmental Information, Washington, D.C. USEPA QA/R-5

SWAMP Quality Assurance Management Plan (SWAMP QMP version 1 dated 12/22//2002 and Draft Version 2 dated 08/09/2006)  
<http://www.swrcb.ca.gov/swamp/qapp.html>

### **III QAPP COMPONENTS**

The U.S. Environmental Protection Agency details the components, content, and format required for a QAPP. Following the guidelines provided by the USEPA, a QAPP must contain specific information regarding four main components:

#### **A. PROJECT MANAGEMENT**

This component addresses basic project management, including the project history and objectives, roles and responsibilities of the participants, and other aspects. These elements ensure that the project has a defined goal, that the participants understand the goal and the approach to be used, and that the planning outputs have been documented.

#### **B. DATA GENERATION AND ACQUISITION**

This component addresses all aspects of project design and implementation. Implementation of these elements ensures that appropriate methods for sampling, measurement and analysis, data collection or generation, data handling, and QC activities are employed and are properly documented.

#### **C. ASSESSMENT AND OVERSIGHT**

This component addresses the activities for assessing the effectiveness of the implementation of the project and associated QA and QC activities. The purpose of the assessment is to provide project oversight that will ensure that the QA Project Plan is implemented as prescribed.

#### **D. DATA VALIDATION AND USABILITY**

This component addresses the QA activities that occur after the data collection, laboratory analysis and data generation phase of the project is completed. Implementation of these elements ensures that the data conform to the specified criteria, thus achieving the project objectives (USEPA 2001).

These four main components are further subdivided into twenty-four (24) specific elements as required by the USEPA. The State SWAMP QAPP guidelines further define items required under each component to ensure that adequate detail is presented within the project's QAPP. The ILRP has additional requirements under each component. In order to provide more information in preparing the QAPP, all required components, elements, and subsections are discussed in the ensuing sections of this document. A QAPP that is submitted for compliance with the ILRP must contain all of the components, elements, and requirements that are described in this document.

### **IV QAPP ELEMENTS**

This section identifies the elements that further describe the four key QAPP components required by the ILRP Program.

#### **A. PROJECT MANAGEMENT**

##### 1 TITLE AND APPROVAL SHEET (USEPA Element 1)

The Title and Approval Sheet element provides the basic project information including the project title, QAPP version number and date, identifies key project staff, and official approval signatures. The Title and Approval Sheet must include the following components:

- 1.1 Project title.**
- 1.2 Revision number.**
- 1.3 Organization name.**
- 1.4 Signature and date block for project lead.**

- 1.5 Signature and date block for project manager(s).**
- 1.6 Signature and date block for project QA officer(s).**

## 2 TABLE OF CONTENTS (USEPA Element 2)

The Table of Contents element provides for organized index of all QAPP components and must include the following components:

- 2.1 List of QAPP sections.**
- 2.2 List of tables and figures.**
- 2.3 List and description of appendices.**
- 2.4 List and description of attached SOPs.**
- 2.5 SOPs revision number and date for each referenced SOP.**

## 3 DISTRIBUTION LIST (USEPA Element 3)

The Distribution List element provides for a comprehensive list of individuals and organizations that will require a copy of the approved QAPP and subsequent revisions. This element also provides for a list of those responsible for implementation of the approved QAPP as well as assessment of compliance of the terms within. The Distribution List element must include the following components:

- 3.1 List of contact staff, organization, phone numbers, email addresses.**
- 3.2 List of names of individuals and organizations that will receive and retain a copy of the QAPP.**

## 4 PROJECT ORGANIZATION (USEPA Element 4)

The Project Organization element provides for a detailed breakdown of key participating individuals and organizations identifying their individual roles and responsibilities within the project. This element also provides information about the chain of authority and at what level key decisions and project assessment reviews will take place. Outside data sources should also be included. The Project Organization element must include the following:

- 4.1 Identify key individuals involved in any major aspect of the project.**
- 4.2 Discuss each individual's responsibility.**
- 4.3 Describe organizational chart detailing lines of authority.**
- 4.4 Designate a QA Manager.**
- 4.5 Identify (if applicable) the individual(s) responsible for maintaining the official, approved QAPP.**
- 4.6 Identify (if applicable) any advisors to the project.**

## 5 PROBLEM DEFINITION/BACKGROUND (USEPA Element 5)

The Problem Definition/Background element provides for a statement of the Project objectives and an overview of historical background for the problem the project is addressing. Existing and applicable regulatory information should also be identified within this section. The Problem Definition/Background element must include the following:

- 5.1 Describe project objectives.**
- 5.2 Describe approaches to meet the objectives.**
- 5.3 Identify applicable regulatory information, applicable criteria, action limits, TMDLs, and Basin Plan objectives.**

**5.4 Describe the decisions to be made, actions to be taken, or outcomes from the information to be obtained.**

**5.5 Describe the project background or historical information for initiating this project.**

The requirements in Sections A.5.4 and A.5.5 need to be placed in the Project's MRP Plan. However, the QAPP should identify the sections and pages where this information can be found in the specific MRP Plan.

## **6 PROJECT DESCRIPTION (USEPA Element 6)**

The Project Description element provides for a summary of all work that is to be performed and the schedule for implementation. This element also provides for a detailed description of the geographical area where sampling is to be performed. The Project Description element must include the following:

**6.1 Detailed summary of work to be performed.**

**6.2 Detailed schedule of major project work benchmarks.**

**6.3 Detailed geographical information.**

**6.4 Photo reconnaissance of the monitoring sites.**

**6.5 Discussion on resource and time constraints.**

Photo reconnaissance of all monitoring sites must be submitted to Central Valley Water Board once a year along with the target GPS coordinates. At a minimum four pictures should be taken and included in the Project report. These pictures should include:

- (a) A general site overview.
- (b) Upstream view.
- (c) Downstream view.
- (d) Entrance to location where the samples will be collected.

## **7 QUALITY OBJECTIVES AND CRITERIA (USEPA Element 7)**

The Quality Objectives (QOs) and Criteria element provides for the QC objectives as well as performance criteria to achieve those objectives. Objectives and criteria for meeting the objectives should be defined at both the sampling design and analytical measurement levels (see Appendices). The analytical measurement levels must meet the requirements defined for a particular method (Appendix A). The completeness criteria (90%) should be calculated and reported with the submittal of each monitoring report (Appendix B). The following tables and definitions must be included within the QOs and Criteria element of the Project's QAPP:

**7.1 Data quality objectives (Appendix B).**

**7.2 Performance criteria goals.**

**7.3 Monitoring parameters table with practical quantitation limits (PQLs) and analytical methods.**

### **7.3.1 Quantitation Limits.**

Laboratories must establish quantitation limits (QLs) that are reported with the analytical results; these may also be called reporting limits. These laboratory QLs must be less than or equal to the PQLs that are identified in the ILRP Monitoring and Reporting Program (MRP) requirements (Appendix A). The laboratories must have documentation to support quantitation at the required levels. Any modification in reported QLs must be identified and discussed in

the laboratory data report. For example, the reported QL for a measurement will change due to sample dilution. The dilution factor, reason for dilution, and other relevant information must be described in the data report.

Laboratories must also report analytical results with measurements equal to or higher than the Method Detection limit (MDL) and lower than the QL. These results must be reported as numerical values and qualified as estimated. Reporting such values as “trace” or “<QL” is not acceptable.

Each laboratory performing analyses for the ILRP program must routinely conduct MDL studies to establish the maximum sensitivity (lowest concentration detectable) for each chemical constituent (Appendix A), and to document that the MDLs are less than the PQLs. The MDL studies must be thoroughly documented and conducted in accordance with Revision 1.1, Code of Federal Regulations (CFR), Title 40, Part 136, Appendix B (1984), “Definition and Procedure for the Determination of the Method Detection Limit.” New MDL studies should be conducted whenever there is a significant change in methods, reagent type or procedures, or within two years of the date the most recent study was conducted.

An MDL is developed from seven aliquots of a standard containing all analytes of interest spiked at approximately five times the expected MDL, which are taken through the analytical method sample processing steps. The data are then evaluated and used to calculate the MDL. If the calculated MDL is less than one-third the spiked concentration, the MDL study must be repeated using a lower concentration.

Project samples may not be analyzed and reported until the MDL study has been completed according to the CFR requirements. MDL study results must be available for review during audits, data review, or as requested. Current MDL study results must be reported at the beginning of every project for review and inclusion in project files.

If any analytes have MDLs that are higher than the project QLs, the following steps must be taken:

- (a) Optimize the sensitivity of the analytical system (as allowed under the appropriate method), and perform a new MDL study sufficient to establish analyte identification at concentrations less than the project-specified QLs.
- (b) If MDLs below required PQLs still could not be achieved for the required constituents using the methods identified in the MRP, the ILRP staff must be contacted. If an alternate method (accredited, modified or performance based) may be used to meet the desired MDLs, a written request to use that method must be provided to the ILRP. The request to use an alternate method must be approved by the Executive Officer and Quality Assurance Officer prior to sample analysis.
- (c) If methods or laboratories that meet the QL requirements are not available, or cannot be feasibly accessed, a variance or exception to a specific QL may be requested in writing. Variances will only be

approved on a case-by-case basis, and after consideration of the impact of the variance, and the documentation provided.

### 7.3.2 Quality control measurements.

The collection of samples and evaluation of data shall provide data that are representative, comparable, complete, precise, and accurate.

*(a) Representativeness:* Sampling locations should be selected that adequately represent all of the discharges from the farm/ranch, or project area, and the affected water bodies. Samples must also be collected during times and at locations that are representative and that meet the objectives described in the ILRP's MRP. Objectives include adherence to sampling Standard Operating Procedures (SOPs), holding times, decontamination procedures, etc.

*(b) Comparability:* Data collected under the ILRP must be comparable in content and quality to the statewide consistency goals outlined by the SWAMP program. An acceptable, approved MRP Plan and project QAPP ensures comparability with other State monitoring programs and projects.

*(c) Completeness:* Data completeness is defined as a measure of the amount of valid data obtained from a measurement system as compared to the planned amount, usually expressed as a percentage. Factors that affect data completeness include sample breakage during transport or handling, insufficient sample volume, laboratory error, QC failure and equipment failure. The dischargers should strive to meet a goal of 90% data completeness per sample batch (Appendix B) and must be calculated and reported with the completion of each monitoring report.

Project completeness can be divided into two areas: Field & Transport Completeness and Laboratory Completeness. Completeness goals should be applied to all aspects within these two areas to meet the 90% total requirement.

Field & Transport Completeness refers to the complete event process of successful planned site visit, conditions documentation, in-field measurements, sample collection technique and volume, in-field quality assurance and control sample preparation, chain-of-custody documentation, preservation, and successful transport of samples to the receiving agencies. Note that if a site is inaccessible or dry, the adequate documentation of these conditions through field sheets, photos, and other means meets the completeness goal for that site and event. Meeting this requirement does not supersede any further requirements outlined in the MRP order that would determine site re-visitation or site location changes.

Laboratory Completeness refers to the complete event process of sample reception, chain-of-custody documentation, storage and in-house preservation, extraction, analysis, and laboratory quality assurance and control samples and measures.

The Project must provide a narrative describing this assessment for each area as well as outline goals for improvement or maintenance of the 90% completeness requirement.

*(d) Precision and Accuracy:* The evaluation of precision and accuracy takes place at the analytical measurement level for values obtained both in the field and in the laboratory. These are further defined in the Appendices of this document, and the calculations to determine the precision and accuracy values are described in Section IV.B.5 of this document.

## **8 SPECIAL TRAINING NEEDS/CERTIFICATION (USEPA Element 8)**

The Special Training Needs/Certification element provides for information regarding any training that will be required for field, laboratory, and other project staff and states the individuals or organizations that are responsible for ensuring that the training is adequate and is completed. The Special Training Needs/Certification element must include the following components:

- 8.1 Identify project personnel with specialized training or certification.**
- 8.2 Identify project field personnel training.**
- 8.3 Identify QA manager and Training Officer.**
- 8.4 Discuss renewal or how new training/certifications will be provided.**
- 8.5 Discuss how training is provided.**
- 8.6 Identify how training is documented.**
- 8.7 Identify the location for staff training records.**

All staff performing field, laboratory, data entry, and data quality assurance procedures shall receive training to ensure that the work is conducted correctly and safely. At a minimum, all staff shall be familiar with the field guidelines and procedures and the laboratory standard operating procedures (SOPs) included in the project QAPP. It is the responsibility of the discharger and project management to ensure that training is mandatory for all personnel, and that such training is documented through training certifications or records. The QA officer for the project is responsible for training but others may conduct training. These records must be maintained and updated for all participating field and laboratory staff.

## **9 DOCUMENTS AND RECORDS (USEPA Element 9)**

The Documents and Records element describes the required documents and records necessary for project quality assurance, including the Project QAPP. The Documents and Records element must include the following components:

- 9.1 Identify reporting format as required by the MRP.**
- 9.2 List all other project documents.**
- 9.3 Discuss where project information will be kept and length of retention.**
- 9.4 Discuss paper and electronic backup methods.**
- 9.5 Discuss how documents will be updated and the responsible party for the update and distribution.**
- 9.6 Discuss how those on the distribution list will receive the most current version of the approved QAPP.**

Copies of field logs, chain-of-custody forms (Section B.3), sample integrity forms for the contract and subcontract laboratories, original preliminary and final laboratory reports,

and electronic media reports must be kept for review by the Central Valley Regional Water Quality Control Board (Central Valley Water Board) ILRP staff. The project field crew must retain original field logs with copies submitted to ILRP staff. The project contract laboratory shall retain original chain-of-custody forms and copies of the preliminary and final data reports for a period of no less than five years.

For each sampling event, the field team or monitoring agency shall provide the Project Lead Staff with copies of the field data sheets, relevant pages of field logs, toxicity laboratory sheets (replicate and in house water quality data) including fail tests, and copies of the chain-of-custody (COC) forms for all samples submitted for analysis. At minimum, the following sample-specific information must be provided for each sampling event:

- (a) Site name.
- (b) Site code.
- (c) GPS coordinates taken with each sampling event.
- (d) Sample type, e.g. grab or composite type (Cross-sectional, flow-proportional, etc.).
- (e) QC sample type and frequency.
- (f) Date and time of sample collection (first sample taken).
- (g) Results of field measurements.
- (h) Sample preservation.
- (i) Requested analyses (specific parameters or method references).
- (j) Results of samples collected and all laboratory QC samples (calibrations, blanks, surrogates, laboratory spikes, matrix spikes, reference materials, etc.) and the identification of each analytical sample batch.
- (k) Results of measurements for tests run prior to toxicity analyses, such as dissolved oxygen, temperature, electrical conductivity, hardness, and ammonia.
- (l) A description of any unusual occurrences, noted by the field personnel, associated with the sampling event - particularly those that may affect sample or data quality.
- (m) Any anomalies regarding sample condition noted by the laboratory.
- (n) Report of any adjustments made to samples prior to running analyses, such as adjustments to dissolved oxygen, alkalinity, de-chlorination, or other.
- (o) Records of exceedance reports or exception reports when results exceed standards or do not meet QC criteria.

For data connectivity purposes all samples taken at a site for one sample event should be assigned one designated sampling time. This time designation is the time assigned to the first sample collected, and must be consistent with the time assigned in the chain of custody, field data sheet, and laboratory report forms. An example of a field data sheet form including all the items described above is included in (Appendix C, Example Form I) at the end of this document.

In the case of field parameters that are continuously monitored through a data logger (e.g. EC, flow, DO, water temperature) field logs are still required as described in items (a) through (n) of this section. The field data should be submitted in the format example provided in Appendix C, Form I. A similar format to the example provided in Appendix C, that contains the required items (see above items (a) through (o)) might be submitted upon Regional Water Quality Control Board approval.

Before measuring field pH a daily check standard is required before the pH measurements are taken. This procedure will help demonstrate that the meter is within acceptable limits.

## **B. DATA GENERATION AND ACQUISITION**

This section describes the elements that are necessary to complete the Data Generation and Acquisition component of the QAPP requirements.

### 1 SAMPLING PROCESS DESIGN (USEPA Element 10)

The Sampling Process Design element provides for discussion on the Project's data collection design in relation to the Project's objectives. This section should include a description of the monitoring approach as well as follow up methods when water quality problems are detected. The Sampling Process Design element must include the following:

- 1.1 Discuss the experimental and data collection design.**
- 1.2 Discuss the rationale for the design.**
- 1.3 Indicate the expected monitoring schedule for each monitoring location.**
- 1.4 Discuss exceedance follow-up plan for each site.**
- 1.5 Indicate the type and total number of samples, matrices, and runs/trials expected or needed for the project.**
- 1.6 Indicate where samples should be taken, and how sites should be identified. A map may be included.**
- 1.7 Describe the course of action should sampling sites become inaccessible.**
- 1.8 Differentiate project data that is critical and data that is for informational purposes only.**
- 1.9 Identify sources of natural variability and how their influence on project data can be minimized.**
- 1.10 Identify potential sources of bias or misrepresentation, and describe how their contribution can be minimized.**

The requirements in Sections B.1.5 through B.1.10 need to be described in the Project MRP Plan. The QAPP must identify the sections and pages where this information can be found in the specific MRP Plan.

### 2 SAMPLE COLLECTION METHODS (USEPA Element 11)

The Sample Collection Methods element provides for information regarding how samples will be collected consistently between all locations and by all sampling staff. The methods for sample collection preparation, physical collection, handling, and transportation must include measures to avoid contamination, ensure accurate tracking, and preserve sample integrity for analysis.

This element also includes a list of applicable field and laboratory Standard Operation Procedures (SOPs) identified by number, date, and regulatory citation. The identified SOPs must be attached to the QAPP as appendixes. Sample Collection Methods element must also include the following components:

- 2.1 Identify criteria for acceptable versus unacceptable water and sediment samples.**

**2.2 Identify pre-sample (Appendices D and E) collection preparation methods.**

**2.3 Identify sample collection method SOPs.**

**2.4 Identify sample container sizes, preservation, and transportation.**

**2.5 Discuss sampling equipment cleansing and decontamination.**

**2.6 Discuss corrective action measures for problematic situations.**

**2.7 Discuss, if applicable to the project, how samples are homogenized, composited, split, and/or filtered.**

**2.8 Describe field procedures including the following items:**

- (a) Photo documentation will occur during all monitoring events as well as GPS coordinates (actual coordinates at the time of sampling). Any changes, in monitoring locations, during monitoring events must be photo-documented and accompanied by GPS coordinates.
- (b) Field personnel must be instructed in the proper collection of samples prior to the sampling event and in how to recognize and avoid potential sources of contamination.
- (c) Field personnel must be able to distinguish acceptable versus unacceptable water and sediment samples in accordance with pre-established criteria.
- (d) Sample containers must be pre-cleaned and certified to be free of contamination according to the USEPA specification for the appropriate methods.
- (e) All field and sampling equipment that will come in contact with field samples must be decontaminated after each use in a designated area to minimize cross-contamination. These details (proper procedures for how and when to clean the equipment) must be specified in the sampling SOP.
- (f) All samples must be identified with a unique number to ensure that results are properly reported and interpreted. Samples must be identified such that the site, sampling location, matrix, sampling equipment, and sample type (i.e., normal field sample or QC sample) can be distinguished by a data reviewer or user.
- (g) A field activity coordinator must be responsible for ensuring that the field sampling team adheres to proper custody and documentation procedures. A master sample logbook or field datasheets shall be maintained for all samples collected during each sampling event.
- (h) All field activities must be adequately and consistently documented to ensure defensibility of any data used for decision-making and to support data interpretation. Pertinent field information, including (as applicable), the width, depth, flow rate of the stream, the surface water condition, location of the tributaries, and the actual GPS coordinates where the sample was taken must be recorded on the field sheets, along with field measurements. All sampling events must include flow information. When possible the USGS method should be used at all wadeable and nonwadeable stream sites for accurately determining flow during each specific monitoring event. If the USGS method cannot be used then flow measurements should be taken near the stream bank of the site or the float method can be used. The approximate location and number of stream flow measurements should be documented on the data sheets. Photo documentation should also be used at all sites for every sample event. Data files for flow data should contain a comment column that will allow a flag for flow measurements that have a high degree of uncertainty. Flow data with a high degree of uncertainty should not be used for pesticide (or other constituent) instantaneous loading

calculations. More rigorous load calculations might be required for TMDL or other programs needs.

### 3 SAMPLE HANDLING AND CUSTODY (USEPA Element 12)

The Sample Handling and Custody element provides for a discussion of the sample integrity maintenance requirements as well as tracking and chain-of-custody procedures. The components of this element must describe the efforts that will be taken to ensure the physical and chemical integrity of a sample from collection to disposal.

Sample Handling Custody element must include the following components:

**3.1 Identify sample holding times, integrity, and storage measures (both before and after extraction). See Appendices D and E for sample handling details.**

**3.2 Identify corrective action for samples that do not meet preservation and/or holding times (Appendix F).**

**3.3 Identify the physical transport of samples from the field.**

**3.4 Discuss sample handling and custody documentation.**

**3.5 Identify sample Chain-of-Custody procedures.**

**3.6 Identify the individuals responsible for verifying procedures.**

**3.7. Describe Field Custody Procedures including the following items:**

- (a) Sample custody must be traceable from the time of sample collection until results are reported. Sample custody procedures provide a mechanism for documenting information related to sample collection and handling.
- (b) A chain-of-custody form must be completed after sample collection and prior to sample shipment or release. The chain-of-custody form, sample labels, and field documentation must be cross checked to verify sample identification, type of analyses, number of containers, sample volume, method of preservation, and type of containers.
- (c) All sample shipments are accompanied with the chain-of-custody form, which identifies the contents. The original chain-of-custody form accompanies the shipment and a copy is retained in the project file.
- (d) All shipping containers must be secured with chain-of-custody seals for transportation to the laboratory. The samples must be transported in ice to maintain sample temperature between 2-4 degrees Celsius. The samples must be sealed in zip lock bags and shipped to the contract laboratories according to Department of Transportation standard.
- (e) Samples that do not meet preservation and/or holding times need to be re-sampled.

#### **3.8. Chain of custody forms**

Chain of custody forms should include the following items:

- (a) Sampler name.
- (b) Address (where the results need to be send).
- (c) Ice chest temperature at log-in.
- (d) To whom the laboratory results need to be sent.
- (e) Laboratory number.
- (f) Field number.
- (g) Lab storage.
- (h) Sample identification.

- (i) Analysis required.
- (j) Number of containers of each type (i.e. plastic, glass, vial, whirlpak).
- (k) Sample collection date and time.
- (l) Comments/special instructions.
- (m) Samples relinquished by (signature, print name, date).
- (n) Samples received by (signature, print name, date).

An example of a Chain of Custody form including all the items described above is attached in the Appendices of this document.

### **3.9. Sample control activities**

Sample control activities must be conducted at the laboratory as well as in the field. Project laboratory custody procedures must include the following conditions:

- (a) Verify initial sample log-in and verification of samples received with the chain-of-custody form.
- (b) Document any discrepancies noted during log-in on the chain-of-custody.
- (c) Initiate internal laboratory custody procedure.
- (d) Verify sample preservation (e.g., temperature).
- (e) Notify the project coordinator if any problems or discrepancies are identified.
- (f) Identify proper sample storage, including daily refrigerator temperature monitoring and sample security.

## **4 ANALYTICAL METHODS AND FIELD MEASUREMENTS** (USEPA Element 13)

The Analytical Methods and Field Measurements element provides for information regarding the specific methods and procedures used to extract, analyze, and/or take measurements of the samples as well as the performance criteria. Analytical Methods and Field Measurements element must include the following components:

- 4.1 Identify methods and SOPs that will meet ILRP requirements.**
- 4.2 Identify instrumentation and kits associated with field measurements and laboratory measurements.**
- 4.3 Describe sample disposal procedures (or refer to Section B.4.1).**
- 4.4 Identify method and instrument performance criteria, detection, and QLs.**
- 4.5 Identify corrective action measures and documentation for test/measurement failure.**
- 4.6 Describe how instruments should store and maintain raw data. Methods or SOPs may be referenced and attached to the QAPP.**
- 4.7 Specify laboratory turnaround times needed.**
- 4.8 Provide method validation and information for all non-standard SOPs and performance based methods (PBMs).**
- 4.9 Indicate where PBMs development records are stored and how they can be accessed.**

If field measurements cannot be collected photo documentation is suggested.

With the inclusion of the above components laboratory analyses discussion in the Project QAPP must also identify the following:

- (a) Laboratory Corrective Actions*

Corrective action measures should also be discussed in the event of instrument failure or performance criteria exceedances. Specific activities that will take place when a failure occurs must be discussed for chemical measurements, toxicity, and microbiological analyses. Project leads must ensure that the laboratory follow the corrective action procedures stated in their QAPP. At a minimum, the approach for corrective action should state the following in the Project QAPP:

“When an out of control situation occurs, analyses or work must be stopped until the problem has been identified and resolved. The analyst responsible must document the problem and its solution and all analyses since the last in control point must be repeated or discarded. The nature and disposition of the problem must be documented in the data report that is sent to the Central Valley Water Board.”

*(b) Laboratory Calibration Curves*

Laboratory adjustments to calibration curves and also to recovery acceptance limits are method dependent. However, when these adjustments are changed during Project implementation, these changes need to be communicated to the ILRP Staff in order to ensure that new limits will meet the Program requirements.

For the ILRP Program, only calibration with a linear regression is acceptable for organic analyses. Non-linear calibration is not allowed due to the fact that using a non-linear option creates a potential for poor quantitation or biased concentrations of compounds at low or high concentrations (near the high and low ends of the calibration range). In order to conduct the linear regression, laboratories shall prepare an initial 5-point calibration curve, where the low level standard concentration is less than or equal to the analyte quantitation limits.

*(c) Pesticide Analyses*

Pesticide analyses must be conducted on unfiltered (whole) fractions of the samples. Prior to the analysis of any environmental samples, the laboratory must have demonstrated the ability to meet the minimum performance requirements for each analytical method. Initial demonstration of laboratory capabilities includes the ability to meet the Project specified quantitation limits (QL), the ability to generate acceptable precision and recoveries, and other analytical and QC parameters as stated in this document.

*(d) Algae Toxicity Testing*

Algae toxicity testing shall not be preceded with treatment of the chelating agent, EDTA. The purpose of omitting this reagent is to ensure that metals used to control algae in the field are not removed from sample aliquots prior to analysis.

*(e) Sediment Toxicity Testing*

The time frame for sediment sample collection, as well as a definition of a "Classified Storm Event" relevant to the project area, shall be described in Section A.6 Project Description of the QAPP. At the time of reporting sediment sample results (exceedance reports and/or SAMR), the project shall also detail the site conditions previous to the sampling event to aid in the analysis of those results. (i.e., details of the last storm in terms of duration and hydrographs or last irrigation details in terms of time, duration, flow and others).

Sediment samples shall be collected using a standardized methodology. Methodology to be used shall be identified and detailed in the Project QAPP Section B.2 Sample Collection Methods. Example protocols can be found in references Section V (USGS Guidelines, 1994).

Sediment samples shall be collected with overlying water present at a collection site, or in the absence of overlying water, when the sediment is moist. Analysis results from sediment samples collected in the absence of overlying water should be flagged as potential outlying data points. Sampling of dry sediment shall not be required, however alternative sampling events should be planned to meet the minimum sample collection requirements as outlined in the MRP.

Sampling conditions shall be documented in the both the field notes and photographs for every successful and non-successful monitoring event (IE including planned events when the site is dry upon arrival). The documentation of field conditions at all attempted events aids the project in meeting completeness goals as outlined by the QAPP as well as establishes a continuous documented history of field conditions for monitoring locations.

*(f) Alternative Analytical Methods*

Analytical methods should be identified by number, date, and regulatory citation. Analytical methods used for chemistry analyses must follow a procedure approved by USEPA or provided in Standard Methods for the Examination of Water and Waste Water 19<sup>th</sup> Edition. When there is a program need to analyze for contaminants that do not have USEPA or Standard Methods procedures, then United States Geological Survey (USGS), American Society of Testing Materials (ASTM), and Association of Official Analytical Chemist (AOAC) methods may be used by accredited laboratories.

If ILRP requirements are provided in the referenced documents, then laboratories may still achieve compliance by submitting a performance-based evaluation of their procedure for the Central Valley Water Board Executive Officer's approval. This will require a peer-reviewed published method or performance-based validation method based upon the protocol described by USEPA "Guide to Methods Flexibility and Approval of USEPA Water Methods" (USEPA, 1996).

Laboratory development of a performance-based method (PBM) validation package and Standard Operating Procedures (SOP) are required when analytes or quantification levels are outside the analyte list or differ by ten times the measurement levels stated in the published method. The validation package must include all data for the "Initial Demonstration of Laboratory Capability," which includes:

1. MDL studies (the analyst shall determine the MDL for each analyte according to the procedure in Code 40 of Federal Regulation (CFR) 136, Appendix B using the apparatus, reagents, and standards that will be used in the practice of this method).
2. Initial precision and recovery (IPR)
3. QC samples, where applicable
4. Linear calibration ranges

*(g) References for Analytical Methods*

The analysis of any material required by this Program shall be performed by a laboratory that has accreditation or certification pursuant to Article 3 (commencing with Section 100825) of Chapter 4 of Part 1 of Division 101 of the Health and Safety Code. General guidance for analytical methods is provided in a list of references in Section V of this document. Specific method modifications may be approved by the Executive Officer of the Central Valley Water Board if sufficient justification is provided.

**5 QUALITY CONTROL (USEPA Element 14)**

The QC element provides information regarding the QC activities that will take place for the Project. Definitions for all quality control samples described here are included in the Appendices to this document. A summary table must be provided, which includes required and optional QC and the frequency. The QC summary table should address all sampling, measurement, and analysis techniques. The following must be included within the QC element of the Project QAPP:

*(a) For Chemical Analyses*

At a minimum, one "QC Set" must be included per analytical method batch per Sampling Event. The minimum required samples for chemical analyses must include:

1. Field blank
2. Field duplicate
3. Matrix spike (MS) and matrix spike duplicate (MSD)
4. Laboratory control spike (LCS) and laboratory control spike duplicate (LCSD)
5. Laboratory blank
6. Laboratory duplicate (MS/MSD or LS/LSD pair may serve this function)

*(b) For Microbiological and Toxicity Analyses*

The minimum required QC samples for microbiological tests must include:

1. Field blank
2. Field duplicate
3. Negative control
4. Positive control

The minimum required QC samples for toxicity tests must include:

1. Field duplicate
2. Negative control
3. Reference toxicant

Optional QC samples that might be utilized by project management include travel blanks, equipment blanks, laboratory duplicates, equipment blank/rinsate samples, and field split samples. Definitions for all quality control samples described here are included in the Appendices to this document.

**5.1 Method blank specifications**

Methods blanks, and all laboratories positive and negative controls for other media and analytes, should be conducted, when necessary (depending on the method), upon initiation of sampling.

Although laboratory blanks are important for all analyses, method blanks for low-level analyses can be conflictive. Improvements in analytical sensitivity have lowered detection limits down to the point where some amount of analyte may be detected in even the cleanest laboratory blanks. In these circumstances, the magnitude of a contaminant found in blanks should be compared to the concentrations found in the samples. **Subtracting method blank results from sample results is not permitted**; however, any blank contamination should be discussed with project management, and must be reported in the monitoring reports that are submitted to the ILRP Staff.

When laboratories obtain detectable concentrations of a specific analyte in the method blanks as part of their laboratory quality control, they need to re-extract and re-analyze in the following circumstances:

*“METALS: If any analyte concentration in the method blank is above the PQL, the lowest concentration of that analyte in the associated samples must be 10 times the method blank concentration. Otherwise, all samples associated with that method blank with the analyte’s concentration less than 10 times the method blank concentration and above the PQL must be re-digested and re-analyzed for that analyte. The sample concentration is not to be corrected for the method blank value.*

*ORGANICS: If any analyte concentration in the method blank is above the PQL, all samples associated with that method blank must be re-extracted and re-analyzed for that analyte. The exception to the above requirement is for common laboratory contaminants such as volatile solvents and phthalates where all samples associated with that method blank, with an analyte concentration less than 10 times the method blank concentration and above the PQL must be re-digested and re-analyzed for that analyte.”*

## 5.2 Matrix spike and spike duplicate specifications

An MS and MSD set must be prepared in the laboratory using sample water collected specifically by the project and be analyzed within the same analytical batch as the original samples. Certified Reference Materials shall be used to prepare MS. After measurement of the MS/ MSD, the Accuracy and Precision must be calculated and noted on the monitoring report and electronic record.

(a) Accuracy of MS Recovery is measured as the percent recovery and provides the accuracy of an analytical test measured against an analyte of known concentration that has been added to an actual field sample. Percent recovery for MS/MSD is calculated as follows:

$$\% \text{ Recovery} = \left( \frac{V_{MS} - V_{Ambient}}{V_{Spike}} \right) \times 100$$

Where:

$V_{MS}$  = is the measured concentration of the spiked sample.

$V_{Ambient}$  = is the measured concentration of the original (unspiked) sample.

$V_{Spike}$  = is the concentration of the spike added.

If the percent recovery for any analyte in the MS or MSD is less than the recommended warning limit, the chromatograms and raw data quantitation

reports must be reviewed. Corrective action that is taken and verification of acceptable instrument response must be included in the cover letter discussion as well.

(b) Precision of the MS/MSD pair is measured as the RPD between two spiked samples and is calculated as follows:

$$RPD = \left| \frac{V_{MS} - V_{MSD}}{Mean} \right| \times 100 \%$$

Where:

*RPD* = is the relative percent difference

$V_{MS}$  = is the measured concentration for the matrix spike.

$V_{MSD}$  = is the measured concentration of the matrix spike duplicate.

*Mean* = is the average of the two concentrations, calculated as follows:

$$Mean = \left[ \frac{(V_{MS} + V_{MSD})}{2} \right]$$

The Data Quality Objective (DQO) for Precision in MS/MSDs is 25% or less. If results for any analytes do not meet this DQO, calculations and instruments must be checked, and the analyst may be required to repeat the analysis to confirm the results. If the results repeatedly fail to meet the objectives indicating inconsistent homogeneity, unusually high concentrations of analytes, or poor laboratory precision, then the laboratory is obligated to:

- Halt the analysis of samples,
- Identify the source of the imprecision, and
- Make corrections where appropriate before proceeding.

If an explanation for a low or high percent recovery value is not discovered, the instrument response may be checked using a calibration standard. Low or high matrix spike recoveries may be a result of matrix interferences and further instrument response checks may not be warranted. An explanation for low or high percent recovery values for MS/MSD results must be discussed in a cover letter accompanying the data package to project management and included in the monitoring report to the Central Valley Water Board.

Failure to meet the designated QOs for MS and MSD is indicative of poor laboratory performance. In this case, the laboratory is obligated to halt the analysis of the samples and to identify the source of the problem and make corrections before proceeding.

### **5.3 Laboratory control spike and spike duplicate specifications**

Laboratory Control Spike (LCS) & Laboratory Control Spike Duplicate (LCSD) provides information on the analytical accuracy, precision, and instrument bias. After measurements of the LCS and LCSD, the Percent Recovery (Accuracy) and Relative Percent Difference (Precision) must be calculated and noted on the report and electronic record.

(a) Accuracy as LCS Recovery is the measured as the test measured against the analyte of known concentration that had been added to laboratory purified water. Recovery for Laboratory Control Spikes is calculated as follows:

$$\% \text{ Recovery} = \left( \frac{V_{LCS}}{V_{Spike}} \right) \times 100$$

Where:

$V_{LCS}$  = is the measured concentration of the spike control sample.

$V_{LCSD}$  = is the concentration resulting from the spike amount added.

If the percent recovery for any analyte in the LCS, LCSD is outside the recommended control limit, the chromatograms and raw data quantitation reports must be reviewed. Corrective action that is taken and verification of acceptable instrument response must be included in the cover letter discussion as well.

(b) Precision of the LCS/LCSD pair is measured as the RPD between two laboratory control samples, and is calculated as follows:

$$RPD = \left| \frac{V_{LCS} - V_{LCSD}}{Mean} \right| \times 100 \%$$

Mean is the average of the results from the two LCS samples, calculated as follows:

$$Mean = \left[ \frac{(V_{LCS} + V_{LCSD})}{2} \right]$$

The Data Quality Objective (DQO) for Precision in LCS/LCSDs is 25% or less. If results for any analytes do not meet this DQO, calculations and instruments must be checked, and the analyst may be required to repeat the analysis to confirm the results. If the results repeatedly fail to meet the objectives indicating inconsistent homogeneity, unusually high concentrations of analytes or poor laboratory precision, then the laboratory is obligated to:

- Halt the analysis of samples,
- Identify the source of the imprecision, and
- Make corrections where appropriate before proceeding.

If an explanation for a low or high percent recovery value is not discovered, the instrument response may be checked using a calibration standard. Low or high matrix spike recoveries may be a result of matrix interferences and further instrument response checks may not be warranted. An explanation for low or high percent recovery values for LS/LSD results must be discussed in a cover letter accompanying the data package to project management and included in the monitoring report to the Central Valley Water Board.

Failure to meet the designated QOs for LS/LSD is indicative of poor laboratory performance. In this case, the laboratory is obligated to halt the analysis of the samples and to identify the source of the problem and make corrections before proceeding.

#### 5.4 Test acceptability criteria for toxicity tests

Decision Step 1: If the Control treatment meets all USEPA Test Acceptability Criteria (TAC), then proceed to statistical analyses for determination of the presence of statistically significant reductions in organism survival or algal growth. For samples that exhibit toxicity, the follow-up requirements in the ILRP MRP must be followed.

Proposed Decision Step 2a: If the control exhibits <90% survival, an acute test of a water sample exhibits 90-100% survival, and the program completeness standard is met (e.g., ≥90% of testing performed successfully to meet ILRP Completeness Objective), the test result should be “flagged” to denote <90% survival in the Control treatment. ILRP completeness must be evaluated with each submittal of Annual or Semi-Annual Monitoring Reports.

If an acute test of a water sample exhibits 90-100% survival, and the program completeness objective for the test is not met, then a re-test of the original sample must be initiated within 24 hours of the observation of a Control treatment with <90% survival.

For the fathead minnow test, the laboratory must take the steps to procure test species within one working day, and the re-test must be initiated within one day of fish being available from a supplier. In all cases, both the original test results and the re-test results must be reported by the Project; the re-test results should be flagged to note that the re-test was initiated outside of the holding time limit. New samples must be collected within five working days of the laboratory identifying a second failure in TAC, if the re-test does not meet USEPA TAC.

Proposed Decision Step 2b: A water sample is not considered toxic if all of the following is true:

- The algal test control does not meet the USEPA TAC for variability (i.e., coefficient of variation >20%), and
- A water sample exhibits an algal cell density that is greater than the algal cell density in the control, and
- The average algal growth in the replicates does not overlap with that in the control (i.e., all test sample replicates exhibit greater algae growth than all control replicates), and
- The Program completeness objective is met.

If the program completeness objective for the test is not met, then a re-test of the original sample must be initiated within 24 hours of the termination of the initial algal test. In all cases, both the original test results and the re-test results must be reported by the Project; the re-test results should be flagged to note that the re-test was initiated outside of the holding time limit. New samples must be collected if the re-test does not meet USEPA TAC.

If an algal test Control treatment does not meet the minimum growth TAC of ≥ 200,000 cells/mL, then a retest of the original sample must be initiated within 24 hours of the termination of the initial algal test. Both the original test results and the re-test results must be reported by the Project; the re-test results should be flagged to note that the re-test was initiated outside of the holding time limit. New samples must be collected within five working days of the laboratory identifying a second failure in TAC, if the re-test does not meet USEPA TAC.

Proposed Decision Step 3: If a Control treatment does not meet USEPA TAC, and the associated ambient water sample(s) have <90% survival (for an acute toxicity test) or the algal growth is less than the Control, then the Regional Board will be notified within 1 business day of the observation of the results in question so that an agreement can be reached regarding how to proceed. At a minimum, re-testing of the original sample within 24 hours of the observed test failure will be required and test results should be “flagged.” For the fathead minnow test, the laboratory must take the steps to procure test species within one working day, and the re-test must be initiated within one day of fish being available from a supplier. If re-testing does not begin within 24 hours, then re-sampling must be conducted within 48 hours of the observed test failure. Re-test results should be flagged to note that the re-test was initiated outside of the holding time limit. New samples must be collected within five working days of the laboratory identifying a second failure in TAC, if the re-test does not meet USEPA TAC.

Note: it is important to recognize that when re-testing a sample beyond the 36-hour holding time prescribed in the test method manual, there is a possibility that toxicity will be reduced or completely gone. In addition, when re-sampling at a site, the new sample does not represent the same conditions under which the original sample was collected (this is particularly important to note when sampling is meant to characterize a specific event such as stormwater runoff).

The reporting of data that do not meet USEPA TAC must also include an assessment from the laboratory as to what may have caused the test control performance issue, the laboratory’s corrective measures to prevent future control failures, a comparison of the data against the USEPA test performance measures, and a comparison of the data against the ILRP required completeness criteria in the Project’s QAPP.

### **5.5 Toxicity procedures - toxicity identification evaluation (TIE)**

Water Column toxicity procedures and triggers for initiating TIEs are described in more detail in Section E.1 of the MRP. At a minimum, Phase I TIE procedures shall be conducted to determine the general class (e.g., metals, non-polar organics, polar organics) of the chemical causing toxicity. Phase II TIEs may also be utilized to confirm and identify specific toxic agents. The TIE report to the Water Board must include a detailed description of the specific TIE procedures that were utilized. Some of the currently known and used TIE procedures are summarized in Appendix G.

### **5.6 Field duplicate specifications**

A field duplicate or field split sample will be collected at the rate of 5% for each analysis (or one set per sampling event, whichever is more frequent). The evaluation of field precision must be addressed in the project QAPP. QAPP acceptance criteria for laboratory precision shall be based only on laboratory-based duplicate samples such as duplicate matrix spikes, blank spikes, laboratory control materials, or certified reference materials. For bacterial analyses, no assessment of field precision is required but laboratories are required to meet methodological precision requirements. Field duplicates with failed results (RPD >25%) do not require re-sampling. However, this data should be flagged and field teams should be notified so that the source of error can be identified and corrective actions taken before the next sampling event.

If a field duplicate result is found to be over the water quality trigger limit an exceedance report must be submitted. Results for field samples and field duplicates must be reported independently and not be averaged for determining an exceedance of water quality trigger limits.

## 6 INSTRUMENT/EQUIPMENT TESTING, INSPECTION AND MAINTENANCE (USEPA Element 15)

The Instrument/Equipment Testing, Inspection and Maintenance element provides for information regarding how personnel can assure that equipment will function properly when needed, as well as the methods for recording equipment failure to track problematic units. The Instrument/Equipment Testing, Inspection, and Maintenance element must include the following components:

- 6.1 Identify field and laboratory equipment that require periodic maintenance and the schedule.**
- 6.2 Identify equipment testing criteria and procedures.**
- 6.3 Identify the individual(s) responsible for instrument/equipment testing, inspection, and maintenance.**
- 6.4 Note the availability and location of spare parts.**
- 6.5 Identify pre-use equipment inspection procedures.**
- 6.6 Identify corrective action measures and documentation for equipment failure.**

## 7 INSTRUMENT/EQUIPMENT CALIBRATION AND FREQUENCY (USEPA Element 16)

The Instrument/Equipment Calibration and Frequency element provides for information regarding how continual quality performance of equipment and instruments will be ensured. The Instrument/Equipment Calibration and Frequency element must include the following components:

- 7.1 Identify field and laboratory equipment that require calibration.**
- 7.2 Identify the calibration procedure and schedule.**
- 7.3 Identify calibration documentation methods.**
- 7.4 Identify corrective action measures and documentation for equipment deficiencies.**

Routine field instrument calibration must be performed at least once per day prior to instrument use to ensure instruments are operating properly and producing accurate and reliable data. Calibration should be performed at a frequency recommended by the manufacturer, if more frequent than once per day and in case of instrument failure. The calibration should be recorded within a field calibration log or directly on the corresponding field sheet.

## 8 INSPECTION/ACCEPTANCE OF SUPPLIES AND CONSUMABLES (USEPA Element 17)

The Inspection/Acceptance of Supplies and Consumables element provides for information regarding how supplies and consumables (e.g., standard materials and solutions, sample bottles, calibration gases, reagents, hoses, DI water, potable water, electronic data storage media) shall be inspected and accepted for use in the project if

applicable. All stock standards and reagents used for extraction and standard solutions must be tracked through the laboratory. The preparation and use of all working standards must be recorded in bound laboratory notebooks that document standards traceable to USEPA, A2 LA or National Institute for Standards and Technology (NIST) criteria.

Records must have sufficient detail to allow determination of the identity, concentration, and viability of the standards including any dilutions performed to obtain the working standard. Date of preparation, analyte or mixture, concentration, name of preparer, lot or cylinder number, and expiration date, if applicable, must be recorded on each working standard. The Inspection/Acceptance of Supplies and Consumables element must include the following components:

- 8.1 Identify critical supplies and consumables for the field and laboratory.**
- 8.2 Identify the source, acceptance criteria, and procedures for the tracking, storing, and retrieving of the above materials.**
- 8.3 Identify the individual responsible for these tasks.**

#### 9 NON-DIRECT MEASUREMENTS (USEPA Element 18)

The Non-Direct Measurements element provides for an identification and discussion of the types of data needed for project implementation or decision making that are obtained from non-measurement sources such as computer data bases, programs, literature files, and historical data bases. The Non-Direct Measurements element must include the following components:

- 9.1 Identify non-direct sources of data that will be used within the project.**
- 9.2 Discuss the intended use of this information.**
- 9.3 Identify the acceptance criteria for the data used.**
- 9.4 Identify any required resources and support facilities (e.g. Data Logger, Controllers).**
- 9.5 Describe the process by which the project determines limits to validity and operating conditions.**

#### 10 DATA MANAGEMENT (USEPA Element 19)

The Data Management element provides for a detailed discussion of the data management process, tracing the path of the data from their generation to their final use and storage.

Data generated shall be converted to a SWAMP comparable format and maintained by the responsible party and available for electronic data submission to the Central Valley Water Board staff. With the inclusion of the above requirement, the Data Management element must include the following components:

- 10.1 Identify the data management scheme from field to final use and storage for all data types.**
- 10.2 Identify standard record keeping and tracking practices and the corresponding SOPs where applicable.**
- 10.3 Discuss how field data and laboratory data will be entered or uploaded into the required data submission format.**

**10.4 Discuss the control mechanism for detecting and correcting errors and for preventing loss of data during data reduction, data reporting, and data entry to forms, reports, and/or database.**

**10.5 Identify the individual(s) responsible for data management.**

**10.6 Verify that continuous monitoring data will be stored in its original Sonde file.**

**10.7 Include any checklists or forms used in data management.**

Procedures for data reduction with respect to significant figures must incorporate the following conventions:

A digit is significant if it is required to express the numerical value of a measurement. The number of significant digits in a measurement must be restricted by the least accurate of its input measurements. These input measurements include all of those associated with sample processing, including aliquots measured during sampling, preparation, and laboratory analysis.

Results of mathematical calculations shall have the same number of significant figures as the calculation's least precise input value. Results of addition and subtraction of measurements shall reflect the decimal position of the calculation's least precise input value. The number of significant figures can vary during these calculations. The final digit in an expressed measurement inherently possesses an uncertainty. This is especially relevant in the discussion of MDLs and reporting limits (RLs). In these instances, the number of reported significant digits must realistically reflect the laboratory's analytical precision.

When the result of a calculation contains too many significant digits, it must be rounded. If a result's trailing digit is less than five, the last significant digit is not changed. If this trailing digit is equal to or greater than five, the last significant digit is rounded up.

## **C. ASSESSMENT AND OVERSIGHT**

### **1 ASSESSMENT AND RESPONSE ACTIONS (USEPA Element 20)**

The Assessments and Response Actions element provides information regarding how a project's activities will be assessed during the project to ensure that the QAPP is being implemented as approved. The Assessments and Response Actions element must include the following:

**1.1 The number, frequency, and type of project assessment activities that will be conducted.**

**1.2 The individual(s) responsible for conducting assessments and indicate their authority to stop work as necessary.**

**1.3 How and to whom assessment information should be reported.**

**1.4 Corrective action measures and documentation for assessment conclusions.**

For existing data use projects, data may be assessed to determine suitability for their intended use and to identify whether project specifications were met. Field operation audits, laboratory performance evaluations, and technical system audits should also be

included in a project's assessment element. The Central Valley Water Board staff may also audit laboratories during sample analyses for this program.

The contractor should routinely observe field operations to ensure consistency and compliance with sampling specifications presented in this document and QAPP that will be developed later. An audit checklist should document field observations and activities.

Performance evaluation (PE) audits quantitatively assess the data produced by a measurement system. Performing an evaluation audit involves submitting certified samples for each analytical method. The matrix standards are selected to reflect the concentration range expected for the sampling program. Any problem associated with PE samples must be evaluated to determine the influence on field samples analyzed during the same time period. The laboratory must provide a written response to any PE sample result deficiencies.

A technical system audit is a quantitative review of a sampling or analytical system. Qualified technical staff members perform audits. The laboratory system audit results are used to review operations and ensure that the technical and documentation procedures provide valid and defensible data.

## 2 REPORTS TO MANAGEMENT (USEPA Element 21)

The Reports to Management element provides for information regarding how management will be kept informed of project oversight, assessment, activities, scheduling, and findings. The Reports to Management element must include the following components:

- 2.1 Identify which project QA status reports will be needed and frequency.**
- 2.2 Identify individual(s) responsible for composing the reports and the individual/s who will receive and respond to the reports.**

The element will identify those responsible for writing reports, when and how often these reports will be written, and identify who will be notified of audit findings. The element will also include the actions project management will take in response to the reports.

## **D. DATA VALIDATION AND USABILITY**

### 1 DATA REVIEW, VERIFICATION AND VALIDATION (USEPA Element 22)

The Data Review, Verification, and Validation element provides the criteria used to review and validate data. These steps help ensure that the data satisfies the quality criteria detailed and required by the ILRP. The Data Review, Verification, and Validation element must include the following:

**ASSESS THE CRITERIA USED TO VALIDATE PROJECT DATA (refer to element A.7)**

Data must be consistently assessed and documented to determine whether project QOs have been met, quantitatively assess data quality, and identify potential limitations on data use. Assessment and compliance with QC procedures should be under-taken throughout the project to ensure the accuracy of sample collection, laboratory analysis, exceedance communications, and the submitted monitoring

reports. Data communicated to Central Valley Water Board staff will be considered draft until the receipt of the monitoring report, which will include copies of signed laboratory data sheets.

The Project QAPP must be used to accept, reject, or qualify the data generated by the laboratory. The Project Manager shall convey the QA/QC acceptance criteria to the laboratory management. The laboratory management will be responsible for validating the data generated by the laboratory. The laboratory personnel must verify that the measurement process was "in control" (i.e., all specified data quality objectives were met or acceptable deviations explained) for each batch of samples before proceeding with analysis of a subsequent batch. In addition, each laboratory will establish a system for detecting and reducing transcription and/or calculation errors prior to reporting data.

The laboratory will submit only data which have met QO's, or which have deviations that are thoroughly evaluated and described, as final results. When QA requirements have not been met, the samples will be reanalyzed when possible and only the results of the reanalysis will be submitted, provided they are acceptable. The Project Manager will be responsible for determining if the validated laboratory data meets the project acceptance criteria.

After data entry or data transfer procedures are completed for each sample event, data should be inspected for data transcription errors, and corrected as appropriate. After the final QA checks for errors are completed, the data should be added to the final database. Quality assurance checks shall be performed at a project level prior to submission within monitoring reports and electronic data submittals.

## 2 VERIFICATION AND VALIDATION METHODS (USEPA Element 23)

The Verification and Validation Methods element provides for the identification of methods or processes for verifying and then validating project information. The Verification and Validation Methods element must include the following components:

- 2.1 Identify the methods and processes used to verify and validate project data.**
- 2.2 Identify the individual(s) responsible for verification and validation of each type of data (e.g., Field Logs, Chain-of-Custodies, Calibration Information, Completeness).**
- 2.3 Identify documentation and or corrective action for discrepancies.**
- 2.4 Attach any checklists, forms, and calculations that will be used.**

The methods to be used or processes to be followed can be identified as SOPs, if available, or described in the text.

## 3 RECONCILIATION WITH USER REQUIREMENTS (USEPA Element 24)

The Reconciliation with User Requirements element provides for a discussion on how validated data will be evaluated to see if it answers the original questions asked within the monitoring objectives. The Reconciliation with User Requirements element must include the following components:

- 3.1 Discuss the procedures to evaluate the uncertainty of the validated data.**
- 3.2 Discuss how limitations on data use should be reported to data users.**

This element outlines the proposed methods to analyze the data and determine possible anomalies or departures from assumptions established in the planning phase of data collection. The element will also describe how reconciliation with user requirements will be documented, issues will be resolved, and how limitations on the use of the data will be reported to decision makers.

## V REFERENCES

Church, P. E., G. E. Granato, and D. W. Owens. 1999. Basic requirements for collecting, documenting, and reporting precipitation and stormwater-flow measurements. United States Geological Survey. Report 99-255, Northborough, Massachusetts.

Guidelines for Collecting and Processing Samples of Stream Bed Sediment for Analysis of Trace Elements and Organic Contaminants for the National Water-Quality Assessment Program; Open-File Report 94-458; U.S. Geological Survey: Sacramento, CA, 1994.

Harrington, J. and M. Born. 2000. Measuring the health of California streams and rivers – A methods manual for water resource professionals, citizen monitors and natural resource students. Report. Sustainable Land Stewardship International Institute, Sacramento, California.

Standard Methods for the Examination of Water and Wastewater, American Public Health Association, American Water Works Association, Water Environment Federation.

SWAMP 2002. Surface Water Ambient Monitoring Program. Quality Assurance Management Plan.

SWAMP 2006. Surface Water Ambient Monitoring Program. Draft Quality Assurance Management Plan.

U. S. Army Corps of Engineers 1981. Procedures for handling and chemical analysis of sediment and water samples. Report. Author: Plumb, R. H. 1981.

USEPA 2001. Laboratory Documentation Requirements for Data Evaluation (R9QA/004.1)

USEPA. 1983. Methods for Chemical Analysis of Water and Wastes. USEPA-600/4-79-020, Third edition.

USEPA. 1988. Methods for Determination of Organic Compounds in Drinking Water (USEPA-600/4-88/039).

USUSEPA. 1998. Methods for Aquatic Toxicity Identification Evaluations. Phase I Toxicity Characterization Procedures. Office of Research and Development, Duluth, MN. USEPA-600-3-88-034.

USEPA. 1998. Methods for Aquatic Toxicity Identification Evaluations. Phase II Toxicity Identification Procedures. Office of Research and Development, Duluth, MN. USEPA-600-3-88-035.

USEPA. 2002. Methods for Measuring the Acute Toxicity of Effluents and Receiving Waters to Freshwater and Marine Organisms, Fifth Edition. Office of Water, Washington, D.C. USEPA-821-R-02-012.

USEPA. 2002. Short-term Methods for Estimating the Chronic Toxicity of Effluents and Receiving Waters to Freshwater Organisms, Fourth Edition. Office of Water, Washington, D.C. USEPA-821-R-02-013.

USEPA. 2001 (2006). USEPA Requirements for Quality Assurance Project Plans (QA/R-5) Office of Environmental Information, Washington, D.C. USEPA QA/R-5.

USEPA. 1994. Methods for Measuring the Toxicity and Bioaccumulation of Sediment-associated Contaminants with Freshwater Invertebrates. Office of Research and Development, Washington, D.C. USEPA-600-R-94-024. Modifications to the procedure for *Hyallela azteca* with respect to the growth endpoint for may be requested in the Project QAPP. TRIGGERS FOCUS GROUP RECOMMENDATION.

USEPA. 1998. Methods for Aquatic Toxicity Identification Evaluations. Phase I Toxicity Characteristics Procedures. Office of Research and Development, Duluth, Minnesota. USEPA-600-3-88-034.

USEPA Test Methods for Evaluating Solid Waste, Physical Chemical Methods, SW 846.

**APPENDIX A: LTMS ANALYTICAL REQUIREMENTS**

Constituents, Parameters, and Tests	Analytical Methods	Reporting Limit	Reporting Unit
Flow	USGS (R2Cross streamflow Method)	1	cfs
pH	SM 4500 H+B, AS 3778 or USEPA 150.1	0.1	pH units
Electrical Conductivity	USEPA 9050A or 120.1	100	µmhos/cm
Dissolved Oxygen	SM 4500-O	0.1	mg/L
Temperature	SM 2550	0.1	° Celsius
Turbidity	SM 2130B or 180.1	1	NTUs
Total Dissolved Solids	SM 2540C or 160.1	10	mg/L
Total Suspended Solids	SM240D or 160.2	10	mg/L
Hardness	USEPA 200.7, 130.1, 130.2, SM 2340C	10	mg/L
Total Organic Carbon	SM 5310C, USEPA 415.1, 415.2	0.5	mg/L
Fecal coliform	SM 9221B/E or 9223	2	MPN/100ml
E-coli	SM 9221B/E (MUG) or 9223	2	MPN/100ml
Algae - <i>Selenastrum capricornutum</i>	USEPA-821-R-02-013	NA	Cell/ml and % Growth
Water Flea - <i>ceriodaphnia</i>	USEPA 821-R-02-012	NA	% Survival
Fathead Minnow - <i>Pimephales promelas</i>			% Survival
Toxicity Identification Evaluation	USEPA-600-3-88-034 and 600-3-88-0355	NA	Stressor Type

Constituents, Parameters, and Tests	Analytical Methods	Reporting Limit	Reporting Unit
<b>Carbamate Pesticides</b>	USEPA 8321 or 632		
Aldicarb	"	0.5	µg/L
Carbaryl	"	0.5	µg/L
Carbofuran	"	0.5	µg/L
Methiocarb	"	0.5	µg/L
Methomyl	"	0.5	µg/L
Oxamyl	"	0.5	µg/L
<b>Organochlorines Pesticides</b>	USEPA 608, 8081A or B, 8272, or 8081		
DDD	"	0.02	µg/L
DDE	"	0.01	µg/L
DDT	"	0.01	µg/L
Dicofol	"	0.1	µg/L
Dieldrin	"	0.01	µg/L
Endrin	"	0.01	µg/L
Methoxychlor	"	0.05	µg/L
<b>Organophosphorus Pesticides</b>	USEPA 8141A, 614, 8321, 625m, or 8270		
Azinphos-methyl	"	0.1	µg/L
Chlorpyrifos	"	0.015	µg/L
Diazinon	"	0.02	µg/L
Dichlorvos	"	0.1	µg/L
Dimethoate	"	0.1	µg/L
Dimeton-s	"	0.1	µg/L
Disulfoton (Disyton)	"	0.05	µg/L
Malathion	"	0.1	µg/L

Constituents, Parameters, and Tests	Analytical Methods	Reporting Limit	Reporting Unit
Methamidophos	"	0.2	µg/L
Methidathion	"	0.1	µg/L
Parathion-methyl	"	0.1	µg/L
Phorate	"	0.2	µg/L
Phosmet	"	0.2	µg/L
<b>Herbicides</b>			
Atrazine	USEPA 619 or 507	0.5	µg/L
Cyanazine	USEPA 619 or 507	0.5	µg/L
Diuron	USEPA 8321 or 632	0.5	µg/L
Glyphosate	USEPA 547	5	µg/L
Linuron	USEPA 8321 or 632	0.5	µg/L
Paraquat dichloride	USEPA 549.1	0.5	µg/L
Simazine	USEPA 619, 8141, 625, 8270C, or 507	0.5	µg/L
Trifluralin	USEPA 8141	0.05	µg/L
<b>Metals</b>			
Arsenic	USEPA 200.7, 200.8, 6020, 1639 or 206.3	1	µg/L
Boron	USEPA 200.7 or 200.8	10	µg/L
Cadmium (total and dissolved)	USEPA 200.7, 200.8, 213.2, 6020, SM 3113, 3113B, or Modified USGS 1996	0.1	µg/L
Copper (total and dissolved)	USEPA 200.7, 200.8, 213.2, 6020, SM 3113, 3113B, or Modified USGS	0.5	µg/L

Constituents, Parameters, and Tests	Analytical Methods	Reporting Limit	Reporting Unit
	1996		
Lead (total and dissolved)	USEPA 200.7, 200.8, 239.2, 6020, 1639, SM 3111B, 3113 or Modified USGS 1966	0.5	µg/L
Nickel (total and dissolved)	USEPA 200.7, 200.8, 249.2, 6020, 1639, or Modified USGS 1996	1	µg/L
Molybdenum	USEPA 200.7, 200.8, 6010, 6020, and 3015A	1	µg/L
Selenium	USEPA 200.7, 200.8, 6020, 270.3, or Modified USGS 1996 0.8, or 270.3	1	µg/L
Zinc (total and dissolved)	USEPA 200.7, 200.8, 289.2, 6020, 1639, SM3113B, or Modified USGS 1996	1	µg/L
<b>Nutrients</b>			
Total Kjeldahl Nitrogen	USEPA 351 or SM 4500-NH <sub>3</sub>	0.5	mg/L
Nitrate plus Nitrite as Nitrogen	USEPA 300, 300.1 351.3, 353.2, or SM 4500	0.05	mg/L
Total Ammonia	USEPA 350 or SM4500 NH <sub>3</sub>	0.1	mg/L
Unionized Ammonia (calculated value)			
Total Phosphorous (as P)	USEPA 365.1, 365.4, or SM 4500-P	0.01	mg/L
Soluble Orthophosphate	USEPA 300.1, 365.1, or	0.01	mg/L

Constituents, Parameters, and Tests	Analytical Methods	Reporting Limit	Reporting Unit
	SM 4500-P		
<b>SEDIMENT SAMPLING</b>			
<b>Sediment Toxicity</b>			
<i>Hyalella Azteca</i>	USEPA 600-R-99-064	NA	% Survival
<b>Pesticides</b>			
	USEPA 1660, 8081 8081A or 8270		
Bifenthrin	“	1.0	ng/g
Cyfluthrin	“	1.0	ng/g
Cypermethrin	“	1.0	ng/g
Esfenvalerate	“	1.0	ng/g
Lambda-Cyhalothrin	“	1.0	ng/g
Permethrin	“	1.0	ng/g
Fenpropathrin	“	1.0	ng/g
Chlorpyrifos	USEPA 8141A, 614, 8321, 625m, or 8270	3.0	ng/g
<b>Other sediment parameters</b>			
TOC	USEPA 415.1, USEPA 9060, Wakley Black, and SW-846	200	mg/kg
Grain Size	ASTM D-422, USEPA 1995, and U. S. Army Corps of Engineers 1981.	1	% sand, % silt, % clay, % gravel

a The method reporting limits (MDLs) and Program Reporting Limits (ILRP RLs) are reasonable goals in terms of laboratory availability and capability, and Project Groups should strive to meet them. If the Project Group contract laboratory proposes alternative methods or RLs, the

proposed alternatives and rationale for the changes must be detailed in the QAPP. Any alternative RL must be approved by the Executive Officer prior to use.

- b. Sampling sites that are selected at waterbodies that are direct tributaries to CWA 303(d) listed waterbodies must be monitored for those listed constituents where they are attributed in the CWA 303(d) list as resulting from agriculture, or if the source is unknown.
- c. The sampling volume submitted to the laboratory shall be of sufficient volume to allow for a TIE, if results show TIE is required.
- d. Assuming 1% organic carbon.
- e. Chloride is only required to be sampled in the areas where the Water Quality Control Plan for the Tulare Lake Basin applies.

APPENDIX B: SUMMARY TABLE OF QUALITY OBJECTIVES AND CRITERIA FOR MEASUREMENT DATA

Group	Parameter	Element 7 Requirements			
		Accuracy	Precision	Recovery	Completeness
Field Testing	Dissolved Oxygen	± 0.5 mg/L	± 0.5 or 10%	NA	90%
	Temperature	± 0.5 °C	± 0.5 or 5%	NA	90%
	Conductivity	± 5 %	± 5%	NA	90%
	pH by Meter	± 0.5 units	± 0.5 or 5%	NA	90%
	Turbidity	± 10% or 0.1%, whichever is greater	± 10% or 0.1 %, whichever is greater	NA	90%
Laboratory Analyses	Conventional Constituents in Water (Additionally see Table II)	Standard Reference Materials (SRM, CRM, PT) within 95% CI stated by provider of material. If not available then with 80% to 120% of true value	Laboratory duplicate, Blind Field duplicate, and MS/MSD ± 25% RPD if Result >10X the MDL. Laboratory duplicate minimum.	Matrix spike 80% - 120% or control limits at ± 3 standard deviations based on actual lab data.	90%
	Synthetic Organic Analytes (including PCBs, PAHs, pesticides)	Standard Reference Materials (SRM, CRM, PT) within 95% CI stated by provider of material. For LCS and LCSD 50% to 150% of true value.	Field duplicate, MS/MSD, and LCS/LCSD ± 25% RPD, if Result > 10X the MDL. Minimum requirements are: field duplicate, MSD, and LCD.	Matrix spike 50% - 150% or control limits at ± 3 standard deviations based on actual lab data.	90%
	Trace metals in water, including mercury	Standard Reference Materials (SRM, CRM, PT) 75% to 125%.	Field duplicate, laboratory duplicate, and MS/MSD ± 25% RPD, if Result >10X the MDL.	Matrix spike 75% - 125%.	90%
	Organic compounds (PCBs, PAHs, pesticides) in sediment and semi-volatiles & volatiles in sediment only	Standard Reference Materials (SRM, CRM, PT) within 95% CI stated by provider of material. If not available then with 50% to 150% of true value	Field duplicate, MS/MSD, and LCS/LCSD ± 25% RPD. Minimum requirements are: field duplicate, MSD, and LCD.	Matrix spike 50% - 150% or control limits at ± 3 standard deviations based on actual lab data.	90%
	Trace metals (including mercury) in sediment	Standard Reference Materials (SRM, CRM, PT) 75% to 125%.	Field duplicate, laboratory duplicate, MS/MSD, and LCS/LCSD ± 25% RPD, if Result > 10 X the MDL except Hg in sediment at ± 35%. Minimum requirements are: field duplicate, MSD, and LCD.	Matrix spike 75% - 125%.	90%
	Total organic carbon in sediment and sediment grain size	CRM within the 95% CI stated by the provider. Laboratory Control Material (LCM) ± 20% to 25% of stated value. No accuracy criteria for grain size.	Duplicate within ± 20% if Result > 10X the MDL	± 25% recovery (75% - 125%)	90%
	Bacteria/ Pathogens	Laboratory positive and negative cultures - proper positive or negative response. Bacterial PT sample --within the stated acceptance criteria.	Rlog within 3.27*mean Rlog (reference is section 9020B of 18th, 19th, or 20th editions of Standard Methods	NA	90%
	Toxicity testing	Meet all performance criteria in method relative to reference toxicant.	Meet all performance criteria in method relative to sample replication.	NA	90%
	Trace Methylmercury in Water	Because no Standard Reference Material for methylmercury in water is available, samples of the tissue SRM DORM-2 are analyzed with the water samples to assess accuracy. Data Quality Objectives are 70-130% of true value.	Field Duplicate or Digestion Duplicate ± 25% RPD, if Result > 10X the MDL. MS/MSD ± 25% RPD	Matrix spike 75% - 130%.	90%

**APPENDIX C: FORM TEMPLATES**  
**EXAMPLE FORM I (a): FIELD DATA SHEET FORM INCLUDING ALL THE MINIMUM ITEMS REQUIRED.**

<b>Irrigated Lands Conditional Waiver Program</b>		<b>Coalition:</b>		<b>Section A</b>		
				Page ___ of ___		
				Date: _____		
Site Name _____	Time for the first _____	<b>GPS Position</b>	Lat (dd.dxxxx)	Long (dd.dxxxx)		
<b>Site Code</b> _____	Sample Taken _____	GPS/DGPS				
Sampling Crew Names _____ <small>(first initial and last name)</small>	Monitoring Event: _____	Target				
Wadeability: yes / no _____	Comments: _____	Actual				
		GPS Model				
<b>FIELD OBSERVATIONS</b>		<b>CIRCLE YOUR OBSERVATIONS</b>			<b>Section B</b>	
Dominant Substrate	Concrete, Cobble, Gravel, Sand, Mud, Other					
Site Odor	None, Sulfides, Sewage, Petroleum, Mixed, Other					
Other Presence	Vascular, Nonvascular, Oily Sheen, Foam, Trash, Other					
Water Odor	None, Sulfides, Sewage, Petroleum, Mixed, Other					
Water Clarity	Clear (see bottom), Cloudy (>4" vis), Murky (<4" vis)					
Water Color	Clear, brown, green, Grey					
Sky Code	Clear, Partly Cloudy, Overcast, Fog, Hazy					
Precipitation	None, Foggy, Drizzle, Rain					
Precipitation (last 24 hrs)	Unknown, <1", >1", None					
<b>PROBE MEASUREMENTS</b>		<b>Section C</b>				
	<b>Flow (cfs)</b>	<b>pH</b>	<b>Electrical Conductivity (uS/cm)</b>	<b>DO (mg/L)</b>	<b>Water Temp (°C)</b>	<b>Turbidity (NTU)</b>
Measurement						
Instrument						
Calibration Date						
<b>SAMPLES TAKEN (# of containers filled)</b>		<b>Section D</b>				
	<b>Physical Parameters (Inorganics) 2 x 1L Plastic*</b>	<b>Total Organic Carbon (TOC) 1 x 40 ml vials*</b>	<b>Nutrients (Inorganics) 1 x 1L Plastic*</b>	<b>Metals (Inorganics) 1 x 60 mL Plastic*</b>	<b>Hardness 1 x 250 mL Plastic*</b>	<b>Pesticides Collected (1 L amber bottles)</b>
Samples						
Duplicate						
Blank						
Matrix Spike						
Total # Containers						
(*) Modified by using the specific characteristics of the containers that are being used				Preserved time and conditions		

**EXAMPLE FORM I (b): FIELD DATA SHEET FORM INCLUDING ALL THE MINIMUM ITEMS REQUIRED.**

<b>Irrigated Lands Conditional Waiver Program</b>		<b>Coalition:</b>	Page ____ of ____ Date: _____				
Site Name _____ Time for the first Sample Taken _____ <b>Site Code</b> _____ Sampling Crew Names _____ Monitoring Event: _____ <small>(first initial and last name)</small> Wadeability: yes / no      Comments: _____			<b>Section A</b>				
<b>GPS Position Same as Water Quality Sample? YES / NO</b>		Lat (dd.ddddd)	Long (dd.ddddd)				
GPS/DGPS							
Target							
Actual							
GPS Model							
<b>FIELD OBSERVATIONS</b>		<b>CIRCLE YOUR OBSERVATIONS</b>					
Sediment Composition		Coarse Sand, Fine Sand, Silt/Clay, Cobble, Gravel, Mixed, Hard Pan Clay, Other					
Sediment Odor		None, Sulfides, Sewage, Petroleum, Mixed, Other					
Other Presence		Vascular, Nonvascular, Oily Sheen, Foam, Trash, Other					
Water Odor		None, Sulfides, Sewage, Petroleum, Mixed, Other					
Water Clarity		Clear (see bottom), Cloudy (>4" vis), Murky (<4" vis)					
Water Color		Clear, brown, green, Grey					
Sky Code		Clear, Partly Cloudy, Overcast, Fog, Hazy					
Precipitation		None, Foggy, Drizzle, Rain					
Precipitation (last 24 hrs)		Unknown, <1", >1", None					
Observed Flow		NA, Dry Waterbody Bed, No Observed Flow, Isolated Pool, 0.1 - 1cfs, 1 - 5 cfs, 5 - 20 cfs, 20 - 50 cfs, 50 - 200 cfs, >200cfs					
<b>SAMPLES TAKEN (# of containers filled)</b>		<b>Section C</b>					
	Toxicity	Pyrethroids	Chlorpyrifos *	TOC	Grain Size		
Samples							
Duplicate							
Matrix Spike	Non Applicable			Non Applicable	Non Applicable		
Total # Containers							
(*)				Preserved time and conditions			





**APPENDIX D: SUMMARY OF SAMPLE CONTAINER, VOLUME, INITIAL PRESERVATION, AND HOLDING TIME RECOMMENDATIONS FOR WATER SAMPLES**

Parameters for Analysis in WATER Samples	Recommended Containers (all containers pre-cleaned)	Typical Sample Volume (ml)	Initial Field Preservation	Maximum Holding Time (analysis must start by end of max)
<b>Conventional Constituents in Water</b>				
Alkalinity	Polyethylene bottles (see NOTE <sup>(1)</sup> below)	100 ml	Cool to 4°C, dark	14 days at 4°C, dark
Chloride (Cl), Sulfate (SO <sub>4</sub> ) and Fluoride (F)	“	300 ml	“	28 days at 4°C, dark
Ortho-phosphate (OPO <sub>4</sub> )	“	150 ml	“	48 hours at 4°C, dark
Nitrate + Nitrite (NO <sub>3</sub> + NO <sub>2</sub> )	“	150 ml	“	48 hours at 4°C, dark
Total Kjeldahl Nitrogen (TKN)	“	600 ml	“	Recommend: 7 days Maximum: 28 days Either one at 4°C, dark
Total Dissolved Solids (TDS)	“	1000 ml	“	7 days at 4°C, dark
Ammonia (NH <sub>3</sub> )	“	500 ml	“	48 hours at 4°C and in the dark or if acidified 28 days at 4°C and in the dark
Total Phosphorus (TPO <sub>4</sub> )	“	300 ml	“	28 days at 4°C, dark
Total Organic Carbon (TOC), Dissolved Organic Carbon (DOC)	“	40 ml (one vial)	“	28 days at 4°C, dark
Total Suspended Solids (TSS)	“	1000 ml (two jars)	“	7 days at 4°C, dark
<b>Trace Metals in Water Samples</b>				
Dissolved Metals (except Dissolved Mercury)	60 ml polyethylene bottle, pre-cleaned in lab using HNO <sub>3</sub>	60 ml (one bottle) if salinity <0.5 ppt 180 ml (three bottles) if	Filter at sample site using 0.45 micron in-line filter, or syringe filter. Cool to 4°C, dark. Acidify in lab, within 24 hrs, using pre-acidified container (ultra-pure HNO <sub>3</sub> for pH<2)	Once sample is filtered and acidified, can store up to 6 months at room temperature
Dissolved Mercury	250 ml glass or Teflon bottle, pre-cleaned in lab using HNO <sub>3</sub>	250 ml (one bottle)	Cool to 4°C, dark. Filter in lab within 48 hours, using bench top Hg filtration apparatus. Acidify in lab within 48 hrs, with pre-tested HCL to 0.5%	Once sample is filtered and acidified, can store up to 6 months at room temperature
Dissolved Methylmercury	250 ml glass or Teflon bottle	250 ml (one bottle)	Cool to 4°C, dark. Filter in lab within 48 hours, using bench top Hg filtration apparatus. Acidify in lab within 48 hrs, with pre-tested HCL to 0.5%.	Once sample is filtered and acidified, can store up to 6 months at room temperature
Total Metals (except Total Mercury)	60 ml polyethylene bottle, pre-cleaned in lab using HNO <sub>3</sub>	60 ml (one bottle) if salinity <0.5 ppt 180 ml (three bottles) if salinity >0.5 ppt	Cool to 4°C, dark. Acidify in lab within 48 hrs, with pre-acidified container (ultra-pure HNO <sub>3</sub> ), for pH<2	Once sample is acidified, can store up to 6 months at room temperature
Total Mercury	250 ml glass or Teflon bottle, pre-cleaned in lab using HNO <sub>3</sub>	250 ml (one bottle)	Cool to 4°C, dark. Acidify in lab within 48 hrs, with pre-tested HCL to 0.5%	Once sample is acidified, can store up to 6 months at room temperature
Methylmercury	250 ml glass or Teflon bottle	250 ml (one bottle)	Cool to 4°C, dark. Filter in lab within 48 hours, with pre-tested HCL to 0.5%	Once sample is filtered and acidified, can store up to 6 months at room temperature
Hardness	200 ml polyethylene or glass bottle	200 ml (one bottle)	Cool to 4°C, dark OR Filter and add 2 ml conc. H <sub>2</sub> SO <sub>4</sub> or HNO <sub>3</sub> to pH < 2; Cool to 4°C, dark	48 hours at 4°C, dark 6 months at 4°C, dark

(1)NOTE:

The volume of water necessary to collect in order to analyze for the above constituents is typically combined in four 1-liter polyethylene bottles, which also allows enough volume for possible re-analysis and for conducting lab spike duplicates. This is possible since the same laboratory is conducting all of the above analyses; otherwise, individual volumes apply.

**APPENDIX D: SUMMARY OF SAMPLE CONTAINER, VOLUME, INITIAL PRESERVATION, AND HOLDING TIME RECOMMENDATIONS FOR WATER SAMPLES**

Parameters for Analysis in WATER Samples	Recommended Containers (all containers pre-cleaned)	Typical Sample Volume (ml)	Initial Field Preservation	Maximum Holding Time (analysis must start by end of max)
<b>Synthetic Organic Compounds in Water Samples</b>				
PESTICIDES & HERBICIDES* <input type="checkbox"/> Organophosphate Pesticides <input type="checkbox"/> Organochlorine Pesticides <input type="checkbox"/> Chlorinated Herbicides	1-L amber glass bottle, with Teflon lid-liner (per each sample type)	1000 ml (one container) *Each sample type requires 1000 ml in a separate container	Cool to 4°C, dark If chlorine is present, add 0.1g sodium thiosulfate	Keep at 4°C, dark, up to 7 days. Extraction must be performed within the 7 days; analysis must be performed within 40 days of extraction
<b>Toxicity Testing Water Samples</b>				
<b>Toxicity in water</b>	Four 2.25 L amber glass bottles (recommended volume 4 gallons)	9000 ml	Cool to 6°C, dark	36 hours at 4°C, dark
<b>Toxicity Testing Water Samples</b>				
<b><i>E. Coli</i></b>	Factory-sealed, pre-sterilized, disposable Whirl-pak® bags or 125 ml sterile plastic (high density polyethylene or polypropylene) container	100 ml volume sufficient for both <i>E. coli</i>	Sodium thiosulfate is pre-added to the containers in the laboratory (chlorine elimination). Cool to 4°C; dark.	STAT: 24 hours at 4°C, dark; lab must be notified well in advance
<b>Fecal Coliform</b>	Factory-sealed, pre-sterilized, disposable Whirl-pak® bags or 125 ml sterile plastic (high density polyethylene or polypropylene) container	100 ml volume sufficient for both fecal <u>and</u> total coliform analyses	Sodium thiosulfate is pre-added to the containers in the laboratory (chlorine elimination). Cool to 4°C; dark	STAT: 24 hours at 4°C, dark; lab must be notified well in advance

**APPENDIX E: SUMMARY OF SAMPLE CONTAINER, VOLUME, INITIAL PRESERVATION, AND HOLDING TIME RECOMMENDATIONS FOR BED SEDIMENT SAMPLES**

Parameters for Analysis in WATER Samples	Recommended Containers	Typical Sample Volume (ml)	Initial Field Preservation	Maximum Holding Time
<b>Bed Sediment Samples</b>				
<b>Synthetic Organic Compounds</b>	250 ml amber glass jar with Teflon lid-liner; Pre-cleaned	500 ml (two jars)	Cool to 4C, dark, up to 48 hours	12 months <sup>(1)</sup> (-20C)
<b>Sediment TOC</b>	125 ml <sup>(2)</sup> clear glass jar; Pre-cleaned	125 ml (one jar)	Cool to 4C, dark, up to 48 hours	12 months <sup>(1)</sup> (-20C)
<b>Sediment Grain Size</b>	125 ml <sup>(2)</sup> clear glass jar; Pre-cleaned	125 ml (one jar)	Cool to 4C, dark, up to 28 days	28 days (4C) <i>Do not freeze</i>
<b>Sediment Toxicity Testing</b>	1-Liter wide-mouth polyethylene jar with Teflon lid-liner; Pre-cleaned	2-Liters (two jars filled completely)	Cool to 4C, dark, up to 14 days	14 days (4C) <i>Do not freeze</i>

(1) Sediment samples for Synthetic Organic Compounds and Sediment TOC analysis can be held at 4C for up to 48 hours (of sample collection), and should be analyzed within this 48 hours period, but can be frozen at any time during the initial 48 hours, for up to 12 months maximum at minus (-) 20C.

(2) Sediment samples for TOC AND grain size analysis can be combined in one 250 ml clear glass jar, and sub-sampled at the laboratory in order to utilize holding time differences for the two analyses. If this is done, the 250 ml combined sediment sample must be refrigerated only (not frozen) at 4C for up to 28 days, during which time the sub-samples must be aliquoted in order to comply with separate storage requirements (as shown above).

APPENDIX F: CORRECTIVE ACTIONS

<b>ILRP CONTROL SAMPLES – ORGANIC COMPOUNDS</b>	
<b>Laboratory Quality Control</b>	<b>Required Corrective Actions for Failures</b>
<b>Calibration Standard</b>	Affected samples and associated quality control must be reanalyzed following successful instrument recalibration.
<b>Continuing Calibration Verification</b>	The analysis must be halted, the problem investigated, and the instrument recalibrated. All samples after the last acceptable continuing calibration verification must be reanalyzed.
<b>Laboratory Blank LAB ROUND TABLE RECOMMENDATION 3.0</b>	If any analyte concentration in the method blank is above the PQL, all samples associated with that method blank must be re-extracted and re-analyzed for that analyte. The exception to the above requirement is for common laboratory contaminants such as volatile solvents and phthalates, where all samples with an analyte concentration less than 10 times the method blank concentration and above the PQL must be re-digested and re-analyzed for that analyte.
<b>Reference Material/LCS/LCSD</b>	Affected samples and associated quality control must be reanalyzed if acceptance criteria are exceeded.
<b>Matrix Spike</b>	Results should be reviewed to evaluate matrix interference. If matrix interference is suspected, and reference material recoveries are acceptable, the matrix spike and the matrix spike duplicate result must be qualified.
<b>Matrix Spike Duplicate</b>	Appropriately spiked results should be compared to the matrix spike and evaluated for matrix interference. If matrix interference is suspected and reference material recoveries are acceptable, the matrix spike duplicate result must be qualified.
<b>Laboratory Duplicate</b>	For duplicates with a heterogeneous matrix and/or ambient levels below the reporting limit, failed results may be qualified. Other failures should be reanalyzed as sample volume allows.
<b>Internal Standard</b>	The instrument must be flushed with rinse blank. If, after flushing, the responses of the internal standards remain unacceptable, the analysis must be terminated and the cause of drift investigated.
<b>Surrogate</b>	If holding times prevent reanalysis, affected results should be qualified. The analytical method or quality assurance project plan must detail procedures for updating surrogate measurement quality objectives.
<b>Field Quality Control</b>	<b>Required Corrective Actions for Failures</b>
<b>Field Duplicate</b>	For duplicates with a heterogeneous matrix and/or ambient levels below the reporting limit, failed results may be qualified. All failures should be communicated to the sampling team so that the source of error can be identified and corrective measures taken before the next sampling event.
<b>Field Blank, Travel Blank, Equipment Blank</b>	If contamination of the field blanks and associated samples is known or suspected, the laboratory should qualify the affected data, and notify the sampling team so that the source of contamination can be identified and corrective measures taken prior to the next sampling event.
<b>Periodic Quality Control</b>	<b>Required Corrective Actions for Failures</b>
<b>Method Detection Limit Study</b>	If results do not meet analytical method requirements and the requirements of 40 CFR Part 136 Appendix B, a new MDL study must be performed before sample analysis begins. Participants wishing to exceed mandated method detection limits or reporting limits must obtain written prior to sample analysis.
<b>Proficiency Test, Intercomparison</b>	Results should be subjected to troubleshooting and/or reanalysis. If allowed by the vendor or referee, results may be resubmitted. To further examine the analytical failure, a follow-up proficiency test or intercomparison study should be completed as soon as possible.

APPENDIX F: CORRECTIVE ACTIONS

<b>ILRP CONTROL SAMPLES – TRACE METALS AND CONVENTIONAL ANALYTES</b>	
<b>Laboratory Quality Control</b>	<b>Required Corrective Actions for Failures</b>
<b>Calibration Standard</b>	Affected samples and associated quality control must be reanalyzed following successful instrument recalibration.
<b>Continuing Calibration Verification</b>	The analysis must be halted, the problem investigated, and the instrument recalibrated. All samples after the last acceptable continuing calibration verification must be reanalyzed.
<b>Laboratory Blank LAB ROUND TABLE RECOMMENDATION 3.0</b>	If any analyte concentration in the method blank is above the PQL, all samples associated with that method blank must be re-extracted and re-analyzed for that analyte. The exception to the above requirement is for common laboratory contaminants such as volatile solvents and phthalates, where all samples with an analyte concentration less than 10 times the method blank concentration and above the PQL must be re-digested and re-analyzed for that analyte. The sample concentration is not to be corrected for the method blank value.
<b>Reference Material/LCS/LCSD</b>	Affected samples and associated quality control must be reanalyzed if acceptance criteria are exceeded.
<b>Matrix Spike</b>	Results should be reviewed to evaluate matrix interference. If matrix interference is suspected, and reference material recoveries are acceptable, the matrix spike and the matrix spike duplicate result must be qualified.
<b>Matrix Spike Duplicate</b>	Appropriately spiked results should be compared to the matrix spike and evaluated for matrix interference. If matrix interference is suspected and reference material recoveries are acceptable, the matrix spike duplicate result must be qualified.
<b>Laboratory Duplicate</b>	For duplicates with a heterogeneous matrix and/or ambient levels below the reporting limit, failed results may be qualified. Other failures should be reanalyzed as sample volume allows.
<b>Internal Standard</b>	The instrument must be flushed with rinse blank. If, after flushing, the responses of the internal standards remain unacceptable, the analysis must be terminated and the cause of drift investigated.
<b>Surrogate</b>	If holding times prevent reanalysis, affected results should be qualified. The analytical method or quality assurance project plan must detail procedures for updating surrogate measurement quality objectives.
<b>Field Quality Control</b>	<b>Required Corrective Actions for Failures</b>
<b>Field Duplicate</b>	For duplicates with a heterogeneous matrix and/or ambient levels below the reporting limit, failed results may be qualified. All failures should be communicated to the sampling team so that the source of error can be identified and corrective measures taken before the next sampling event.
<b>Field Blank, Travel Blank, Equipment Blank</b>	If contamination of the field blanks and associated samples is known or suspected, the laboratory should qualify the affected data, and notify the sampling team so that the source of contamination can be identified and corrective measures taken prior to the next sampling event.
<b>Periodic Quality Control</b>	<b>Required Corrective Actions for Failures</b>
<b>Method Detection Limit Study</b>	If results do not meet analytical method requirements and the requirements of 40 CFR Part 136 Appendix B, a new MDL study must be performed before sample analysis begins. Participants wishing to exceed mandated method detection limits or reporting limits must obtain written prior to sample analysis.
<b>Proficiency Test, Intercomparison</b>	Results should be subjected to troubleshooting and/or reanalysis. If allowed by the vendor or referee, results may be resubmitted. To further examine the analytical failure, a follow-up proficiency test or intercomparison study should be completed as soon as possible.

**APPENDIX F: CORRECTIVE ACTIONS**

<b>ILRP CONTROL SAMPLES – FIELD PARAMETERS</b>	
<b>Field Measurement</b>	<b>Required Corrective Actions for Failures</b>
<b>Depth, Dissolved Oxygen, pH, Salinity, Specific Conductance, Temperature, Turbidity, Velocity</b>	The instrument should be recalibrated following its manufacturer's cleaning and maintenance procedures. If measurements continue to fail measurement quality objectives, affected data should not be reported and the instrument should be returned to the manufacturer for maintenance. All troubleshooting and corrective actions should be recorded in the calibration and field data logbooks
<b>ILRP CONTROL SAMPLES – TOXCITY TESTING</b>	
<b>Negative Controls</b>	<b>Required Corrective Actions for Failures</b>
<b>Laboratory Control Water</b>	See Toxicity Trigger's Focus Group Recommendation 8
<b>Conductivity Control Water</b>	Flag the data for samples with similar electrical conductivities (EC) and for the EC control and ensure that EC was within the species tolerance range.
<b>Additional Control Water (Method Blank)</b>	Flag the data for samples affected or compared to the failed method blanks.
<b>Positive Controls</b>	<b>Required Corrective Actions for Failures</b>
<b>Reference Toxicant Tests</b>	Immediately re-set up within 48 hours of failure and investigate source of failure.
<b>Field Quality Control</b>	<b>Required Corrective Actions for Failures</b>
<b>Field Duplicate</b>	Flag the data for samples affected and the source of the failure should be identified to prevent future failures. All QC failures should be reported immediately. If QC samples do not meet completeness criteria the data will be flagged

**APPENDIX G: TOXICITY EVALUATION IDENTIFICATION PROCEDURES**

Phase I Procedures	<i>Ceriodaphnia</i>	<i>Selenastrum</i>	<i>Pimephales</i>	Purpose of Procedure
<b>Addition of piperonyl butoxide</b>	X	NA	NA	Inactivates metabolically activated organophosphorous compounds. Increases toxicity of pyrethroids insecticides.
<b>Aeration</b>	X	X	X	Remove volatile chemicals, surfactants and sublutable compounds.
<b>AG2-X8 Solid Phase Extraction (SPE)</b>	X	X	X	Remove multivalent anions.
<b>Antibiotic Amendment</b>	X	Unknown	X	Reduces pathogen infections.
<b>C8 (C18) SPE</b>	X	X	X	Removes non-polar organic chemicals
<b>C 8 SPE eluate add-back</b>	X	X	X	Confirms presence of non-polar organic compound (s).
<b>Centrifugation</b>	X	X	X	Removes particle-bound chemical and biological contaminants.
<b>Chelation (addition of EDTA)</b>	X	X	X	Inactivates cationic metals (Al, Cd, Cu, Zn, Pb, Fe, Ni).
<b>Chelex SPE</b>	X	X	X	Remove multivalent cations.
<b>Filtration</b>	X	NA	X	Removes particle-bound chemicals and biological contaminants.
<b>Graduated pH adjustment</b>	X	NA	X	Increased pH. Increases ammonia toxicity.
<b>Hardness manipulation</b>	X	Unknown	X	Decreases solubility/speciation of metals (bioavailability).
<b>Oxidation Reduction (addition of sodium thiosulfate)</b>	X	Unknown	X	Inactivates Cu, Se, Ag,Hg, Cd, Mn ions, Br, I, O <sub>3</sub> (Ozone).
<b>Temporary pH shift to 3</b>	X	Unknown	X	Breaks down hydrolyzable organic compounds, may increase metal solubility/speciation (bioavailability).
<b>Temporary pH shift to 11</b>	X	X	X	Precipitates metals (may decrease metal bioavailability). Breaks down hydrolyzable organic compounds.
<b>Ultraviolet Light</b>	X	Unknown	X	Activates polyaromatic hydrocarbons, inactivates biological contaminants.
<b>Zeolite</b>	Unknown	X	X	Removes unionized ammonia
Phase II Procedures	<i>Ceriodaphnia</i>	<i>Selenastrum</i>	<i>Pimephales</i>	Purpose of Procedure
Solvent fractionation of SPE eluate	X	X	X	Identifies specific non-polar organic compounds causing toxicity.
Phase III Procedures	<i>Ceriodaphnia</i>	<i>Selenastrum</i>	<i>Pimephales</i>	Purpose of Procedure
Side-by- side dilution series	X	X	X	Determines the contribution of suspected chemical (s) to toxicity.

NA = Manipulation not compatible for series X = manipulation compatible for series

## APPENDIX H: ONLINE RESOURCES

### Hosted by the State Water Resources Control Board

SWAMP Quality Assurance Management Plan:

<http://www.waterboards.ca.gov/swamp/qamp.html>

*This QAMP and associated appendices in Adobe PDF and Microsoft Word formats*

SWAMP Quality Assurance Project Plan Template:

[http://www.waterboards.ca.gov/swamp/docs/swampqapp\\_template032404.doc](http://www.waterboards.ca.gov/swamp/docs/swampqapp_template032404.doc)

*Template for SWAMP-comparable QAPP creation*

SWAMP Quality Assurance and Quality Control:

<http://www.waterboards.ca.gov/swamp/qapp.html>

*SWAMP quality assurance homepage and links*

### Hosted by the Moss Landing Marine Laboratories

SWAMP Standard Operating Procedures:

<http://mpsi.mlml.calstate.edu/swsops.htm>

*SWAMP data management and quality assurance SOPs*

SWAMP Quality Assurance Comparability:

<http://mpsi.mlml.calstate.edu/swqacompare.htm>

*Guidelines and links pertaining to SWAMP quality assurance comparability*

SWAMP Data Management Comparability:

<http://mpsi.mlml.calstate.edu/swdbcompare.htm>

*Guidelines and links pertaining to SWAMP data management comparability*

**ATTACHMENT D**

**CALIFORNIA REGIONAL WATER QUALITY CONTROL BOARD  
CENTRAL VALLEY REGION**

**MONITORING AND REPORTING PROGRAM  
ORDER NO. R5-2009-00809**

**MONITORING SITE LOCATIONS  
FOR  
CALIFORNIA RICE COMMISSION  
FOR DISCHARGES FROM IRRIGATED LANDS**

The following sites will be monitored according to the schedule and for the parameters listed in this MRP Order.

**CRC 2009 MONITORING SITES**

<b>Site Code</b>	<b>Site Name</b>	<b>Latitude</b>	<b>Longitude</b>	<b>Site Type</b>
CBD1	Colusa Basin Drain above Knights Landing	38.8125 N	-121.7731 W	Core Monitoring
CBD5	Colusa Basin Drain #5	39.1833 N	-122.0500 W	Core Monitoring
BS1	Butte Slough at Lower Pass Road	39.1875 N	-121.9000 W	Core Monitoring
SSB	Sacramento Slough Bridge near Karnak	38.7850 N	-121.6533 W	Core Monitoring
F	Lurline Creek; upstream site of CBD5	39.2184 N	-122.1511 W	Assessment
G	Cherokee Canal, upstream site for BS1*	39.3611N	-121.8675W	Assessment
H	Obanion Outfall at DWR PP on Obanion Rd	39.0258N	-121.7272W	Assessment

\* If there is no flow at the specified site, a site on Butte Slough will be sampled.